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Growth Enhancement of Tomato by a Plant Growth Promoting Bacterium, *Bacillus subtilis* PE7

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ABSTRACT

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Kil Yong Kim https://orcid.org/0000-0002-1757-2908 *Bacillus* species are well recognized as effective bio-stimulants to enhance crop quality and productivity. Our study demonstrates the effect of a plant growth promoting bacterium, *Bacillus subtilis* PE7 on tomato growth. *In vitro* screening assay for plant growth promoting traits indicated that the indole-3-acetic acid (IAA) production of *B. subtilis* PE7 in terms of increasing intensity of pink color with the increasing concentration of L-tryptophan supplemented in tryptone soy broth (TSB) medium. The supplementation of L-tryptophan (1 g L^{-1}) in the TSB medium resulted in the maximum IAA production of strain PE7 with 25.59 µg mL⁻¹ at 6 days after incubation. Moreover, the highest ammonia production of strain PE7 with 21.22 µg mL⁻¹ was observed at an incubation period of 72 h. The results of our pot experiment revealed that treatment with the strain PE7 culture (inoculated in fertilizer-based medium) induced a significant growth promotion of tomato plants resulting in higher values of leaf number, length, fresh and dry weights of shoot as compared to the control and fertilizer treatment. Therefore, *B. subtilis* PE7 could be considered as an effective candidate for the development of eco-friendly commercial fertilizer in crop production.

Keywords: Ammonia production, Bio-stimulant, IAA production, PGPB



Growth promotion of tomato plants by *Bacillus subtilis* PE7 at 5 weeks after transplantation. *Con: water as control, F: fertilizer treatment, PE7: *B. subtilis* PE7 culture.



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Introduction

Tomato (Solanum lycopersicum L.), a member of the Solanaceae family is the second most important vegetable crop after potato due to its economic significance with a production of 189.1 million tons on the area of 5.2 million hectares worldwide (FAOSTAT, 2021). Moreover, tomatoes are natural origins of health-promoting compounds including vitamin B, lycopene, carotenoids, ascorbic acid, and polyphenolic compounds which are representative of anti-tumor, anti-oxidant and anti-inflammatory activities (Quinet et al., 2019). However, the demand for improvement of fruit quality and large comsumption of tomatoes lead to the over-application of synthetic fertilizers contributing to deleterious effects on ecosystems including soil acidification, groundwater contamination and an increase in greenhouse gas level. As a consequence, research has been focused on sustainable agricultural methods through applications of different kinds of biofertilizers which are usually based on plant growth promoting bacteria (PGPB). PGPB including species of Bacillus, Pseudomonas, Enterobacter, Klebsiella, Azotobacter, Clostridium, Azospirillium, and Serratia have been studied and utilized as biofertilizers for decades (Esitken et al., 2010; Minut et al., 2022). Among these strains, *Bacillus* species become more attractive due to their endurance by developing stress-resistant endospores under unfavorable environmental conditions leading to the successful field performance. Like other PGPB, Bacillus species assist plant growth directly by facilitating nutrient efficiency via production of ammonia (NH₃), solubilizing insoluble phosphate and releasing phytohormones such as indole-3-acetic-acid (IAA) and gibberellic acid (GA3) (Glick, 2012; Gohil et al., 2022). Indirectly, they reduce the degree of plant stress hormone, ethylene through production of 1-aminocyclopropane-1-carboxylate (ACC) deaminase and induce the plant defense system by elevating the level of the defense-related compounds in response to pathogen attacks (Glick, 2012; Shin et al., 2016). Moreover, these species also act as biocontrol agents and play a significant role in inhibition of phytopathogens via secretion of diverse active compounds including antibiotics, volatile organic compounds (VOCs), hydrogen cyanide (HCN), and siderophores (Olanrewaju et al., 2017; Lee et al., 2021; Hwang et al., 2022; Kim et al., 2022).

Generally, the group of *Bacillus* species are ubiquitous in a broad-spectrum of environments such as air, soil, water and rhizosphere, and they are also common habitants of fermented foods such as yogurt, cheonggukjang (extra-fermented bean paste from Korea) and kimchi (Lee et al., 2013; Kimura and Yokoyama, 2019; Luo et al., 2023). Kimchi, a traditional fermented Korean side-dish prepared with Chinese cabbage, red pepper, radish and garlic, is a resource of beneficial microbes involving *Bacillus* species. In many reports, the bacterial species isolated from kimchi possessed unique properties with stability at a wide range of pH and temperature (Lee et al., 2013; Radhakrishnan and Lee, 2016; Jeon et al., 2017; Han et al., 2023).

The hypothesis was that the broth culture of *Bacillus subtilis* PE7 isolated from kimchi will promote the growth of tomato as well. In our study, the bacterial strain, *B. subtilis* PE7 was used to evaluate its plant growth promoting (PGP) potential on tomato plants. Therefore, the objectives of this study were to determine ammonia and IAA production by strain PE7 under *in vitro* conditions and to evaluate the effect of the strain PE7 culture on the growth of tomato via pot experiment.

Materials and Methods

Bacterial strain and culture conditions *B. subtilis* PE7 (KACC 92549P) was isolated from kimchi in our previous study (Han et al., 2023), and this bacterial strain was used for the present study. For *in vitro* experiments, ten microliters of original stock culture of *B. subtilis* PE7 was streaked on tryptone soy agar (TSA) medium to attain pure and single colonies. For pot experiment, strain PE7 was inoculated in pink-fertilizer broth (PFB) medium and incubated in a shaking incubator at 40°C for 2 days. One liter of PFB medium consists of dextrose 2.0 g; chitin powder 0.4 g; commercial fertilizer (NPK: 20-20-20) 5.6 g; CaCO₃ 0.3 g; K₂SO₄ 0.4 g, MgCl₂ 0.2 g, and 2 mL of micronutrients.

IAA production For screening of IAA production, *B. subtilis* PE7 was inoculated in tryptone soy broth (TSB) medium containing different concentrations of L-tryptophan (0, 0.25, 0.5 and 1 g L⁻¹) and incubated in a shaking incubator at 30°C for 3 days. The broth cultures were centrifuged at 12,000 rpm and 4°C for 15 min. The supernatant (1 mL) was mixed with of Salkowski's reagent (2 mL) in a test tube and the tubes were placed in the dark at room temperature for 30 min. The development of pink color indicates IAA secretion by *B. subtilis* PE7.

IAA production by *B. subtilis* PE7 was evaluated quantitatively using a UV spectrophotometer (Gusmiaty and Payangan, 2019). The pre-culture of strain PE7 was inoculated in a flask containing TSB medium added with 1 g L^{-1} of L-tryptophan and incubated in a shaker at 30°C and 130 rpm for 7 days. The broth culture was collected every day and centrifuged at 12,000 rpm and 4°C for 15 min. The resulting supernatant (1 mL) was mixed with Salkowski's reagent (2 mL) and incubated in the dark at room temperature for 30 min. After incubation, the absorbance was read at 530 nm by a UV-spectrophotometer and the amount of IAA secreted by *B. subtilis* PE7 was calculated based on the standard curve of authentic IAA.

Ammonia production Ammonia production of *B. subtilis* PE7 was determined following the method by Karthik et al. (2017). A single colony of strain PE7 was pre-inoculated in a flask containing 100 mL of TSB medium at 30°C for 2 days. The pre-inoculated broth culture of strain PE7 was grown in the peptone water and incubated at 30°C for 72 h. Triplicate flasks were used for the experiment. After incubation, the broth culture was collected 12 h intervals and the supernatants were obtained by centrifuging at 12,000 rpm and 4°C for 15 min. One milliliter of each resulting supernatant was mixed with 1 mL of Nessler's reagent. The total volume was increased to 10 mL with double distilled water (DDH₂O) and thoroughly mixed using a vortex mixer. Then, ammonia production by strain PE7 was measured at a wavelength of 450 nm using a UV-spectrophotometer.

Pot experiment Two-leaf stage of tomato seedlings were obtained by sowing the seeds in a tray which contained organic bed soil, and watered every day for 2 weeks. The seedling was transplanted into the pot containing a mixture of soil, sand, and vermiculite (1:1:1, v/v/v, pH: 6.14 ± 0.11 , electrical conductivity (EC): 0.57 ± 0.01 dS m⁻¹). The experiment was designed with three treatments: (1) water as the control (Con), (2) PFB medium as the fertilizer treatment (F), and (3) *B. subtilis* PE7 broth culture (PE7). The experiment was conducted with three replications and five plants were used for each replication. At 7 days after transplantation, 50 mL of each treatment per week was consecutively applied to tomato plants via soil drenching twice for the first and second time-treatment and 100 mL was used twice for third and fourth time-treatment. At 3 days after the final treatment, tomato plants were carefully uprooted and rinsed gently with running tap water. The tomato growth was analyzed based on leaf number, length, fresh weight, and dry weight of the shoot and root. The shoots and roots were oven-dried at 70°C for 3 days and determined the dry weights.

Statistical analysis The data of plant growth parameters were used to analyze by analysis of variance (ANNOVA) using statistical analysis system (SAS) software (9.4). The mean comparison was done by the least significant difference (LSD) test at p < 0.05 level.

Results and Discussion

Qualitative and quantitative determination of IAA production by *B. subtilis* PE7 Several studies reported that *Bacillus* species produced different types of phytohormones including IAA to regulate plant growth (Cherif-Silini et al., 2016; Ratnaningsih et al., 2023). As shown in Fig. 1, the intensity of pink colorization indicated that IAA production of *B. subtilis* PE7 was L-tryptophan-concentration dependent. Moreover, the quantitative assay revealed that IAA production gradually increased from 1 day after incubation (DAI) to 6 DAI in the L-tryptophan supplemented medium, the IAA amount attained the maximum with 25.59 μ g mL⁻¹ at 6 DAI, and then declined at 7 DAI (Fig. 2). In a previous study by Wagi and Ahmed (2019), the presence of L-tryptophan in the Luria-Bertani (LB) medium induced auxin production (35.8 and 36.6 μ g mL⁻¹) by both *Bacillus cereus* (So3II) and *B. subtilis* (Mt3b). Cherif-Silini et al. (2016) also reported that 10.35 μ g mL⁻¹ of IAA was produced by *B. subtilis* subsp. *subtilis* D7 in DF salt minimal medium containing 1 g L⁻¹ of L-tryptophan.

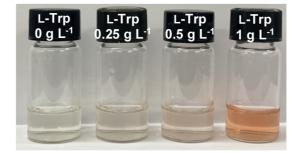


Fig. 1. Intensity of pink color indicates IAA production of *Bacillus subtilis* PE7 affected by different concentrations (0, 0.25, 0.5, 1 g L⁻¹) of L-tryptophan (L-Trp) in the growth medium.

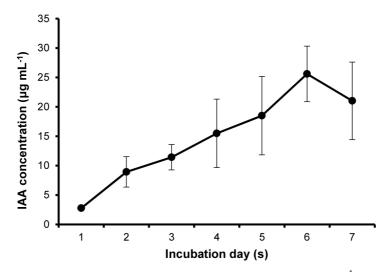


Fig. 2. Indole-3-acetic-acid (IAA) production of *Bacillus subtilis* PE7 in L-tryptophan (1 g L⁻¹) supplemented TSB medium for 7 days.

Ammonia production by *B. subtilis* PE7 Members of *Bacillus* species release ammonia not only to enhance plant growth by supplying the required nitrogen source to host plant but also to protect the plant from the attack of phytopathogens by limiting their growth (Alotaibi et al., 2022; Gohil et al., 2022). Nithyapriya et al. (2021) reported that *B. subtilis* LSBS2 produced 8 μ g mL⁻¹ of ammonia in peptone water after 48 h incubation period. The colori-

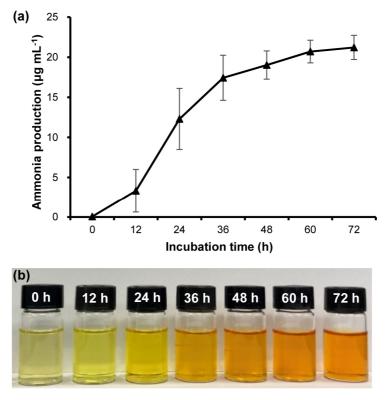


Fig. 3. Ammonia production of *Bacillus subtilis* PE7 in peptone water for 72 h (a) and the degree of ammonia production by *B. subtilis* PE7 with the appearance of yellow to brown color depending on the incubation time (b).

metric assay showed that a gradual increase in ammonia production of strain PE7 within a range of 0.06 to $21.22 \,\mu g$ mL⁻¹ was recorded during the experimental period in the present study. (Fig. 3).

Effect of B. subtilis PE7 culture on growth of tomato The potential of PGPB for plant growth enhancement through direct and indirect regulation has been well documented in several reports (Katsenios et al., 2021; Zhou et al., 2021; Kouam et al., 2023). However, one of the major challenges for field application is expensive growth medium for inoculation of PGPB. Therefore, to overcome the limitation on wide application of bacterial agents for crop production, cheap fertilizer-based PFB medium was used to inoculate B. subtilis PE7 in our study. Our study demonstrated the PGP ability of B. subtilis PE7 on growth of tomato via pot experiment. As shown in Table 1 and Fig. 4, the inoculation of strain PE7 broth culture to tomato plants remarkably improved tomato growth in terms of leaf number, shoot length, fresh and dry weights of shoot as compared to the Con and F treatment. Although, there was no statistical difference of fresh and dry weights of root between PE7 and F treatments, the values of these parameters in PE7 treatment were higher than those of F treatment. The strain PE7 culture remarkably affected tomato growth resulting in a significant increase in leaf number, length, fresh and dry weights of shoot respectively as compared to the control. Although the presence of IAA and ammonia in PFB medium was not verified in this study, the growth promotion of tomato plants could be related with the action of various metabolites which accumulated in the broth culture of strain PE7. Moreover, the viable cells of strain PE7 in the broth culture might also partially involve in growth promotion of tomato plants because our in vitro IAA and ammonia assays revealed that strain PE7 possessed the capacity to produce IAA and ammonia. Bacterial IAA plays a major role in root stimulation, cell division and enlargement, and enhances root development including root elongation and abundant formation of root hairs, thereby improving water and nutrient acquisition by plant root (Bessai et al., 2022; Castillo-Alfonso et al., 2022). In addition, ammonia released by PGPB induces a direct stimulation of plant growth through nitrogen accumulation for shoot and root growth, and biomass production (Kouam et al., 2023). Various PGP properties of strain PE7 such as phosphate and zinc solubilization, siderophore and exopolysaccharide production, have been reported in our previous study (Han et al., 2023). Our results suggest that the simultaneous action of viable cells of strain PE7 and active metabolites in the broth culture were mainly responsible for achieving higher growth parameters of tomato. Ahmed et al. (2022) reported the dual effects of Bacillus amyloliquefaciens WS-10 broth culture grown in LB medium on reduction in disease incidence of tobacco bacterial wilt and an increase in dry matter contents of roots, stems, and leaves compared to the control. Moreover, the use of 100-fold-diluted broth cultures of B. subtilis and B. amyloliquefaciens which were inoculated in TSB medium, increased fruit weight of tomato plant by 30.70% and 28.81%, as compared to the control under field conditions (Katsenios et al., 2021). In a report by Qiao et al. (2017), seed treatment with the cell suspension of B. subtilis PTS-394 between OD₆₀₀ value of 0.01 and 1.0 ($\sim 5 \times 10^7$ CFU mL⁻¹), increased the fresh weight of tomato seedlings in MS agar medium. Among the treatments, the inoculation of cell suspension (~5 \times 10⁷ CFU mL⁻¹) resulted in the greater value of fresh weight of tomato seedlings (18.36%) compared to the control. Similar growth enhancement of tomato plants by the applied cell suspension of *B. subtilis* PTS-394 was also demonstrated in their pot experiment. Also, in our pot experiment, the viable cells of strain PE7 (which have the capacities of IAA and ammonia production) might activate in the soil, thereby promoting tomato plants. Therefore, the viable cells of strain PE7 and its accumulated active metabolites could be considered as eco-friendly effective substitutes for chemical fertilizer to not only enhance tomato growth but also reduce the frequency of fertilizer application in sustainable agriculture.

			5		•		
Treat-	Length (cm)		Fresh weight (g)		Dry weight (g)		- Leaf number
ment [†]	Shoot	Root	Shoot	Root	Shoot	Root	Lear number
Con	$9.40\pm0.28~c^\dagger$	19.63 ± 5.89 a	$2.77\pm0.08\ c$	$0.75\pm0.07\;b$	$0.26\pm0.00\;c$	$0.07\pm0.01\ b$	$6.00\pm0.00\ c$
F	25.03 ± 1.34 b	16.77 ± 2.54 a	$18.22\pm0.88~b$	1.52 ± 0.26 a	$1.10\pm0.08\ b$	0.12 ± 0.03 a	$9.00\pm0.00\ b$

Table 1. Effect of Bacillus subtilis PE7 culture on growth characteristics of tomato plants.

⁺Con: water as control, F: fertilizer treatment, PE7: *B. subtilis* PE7 culture.

 30.30 ± 0.83 a 19.37 ± 3.71 a 29.25 ± 2.23 a 1.79 ± 0.12 a

[†]Different letters in the same column indicate significant difference between treatments as determined by the least significant difference (LSD) test at p < 0.05 level. Data represents mean ± standard deviation of three replicates.

 1.67 ± 0.12 a

 0.13 ± 0.01 a

 11.00 ± 0.00 a

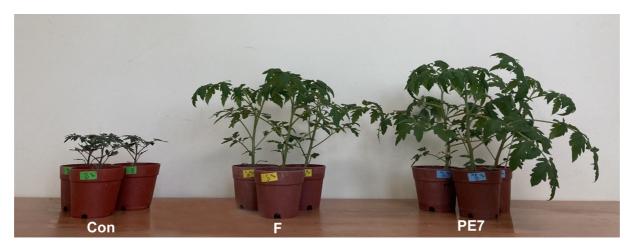


Fig. 4. Growth promotion of tomato plants by *Bacillus subtilis* PE7 at 5 weeks after transplantation. *Con: water as control, F: fertilizer treatment, PE7: *B. subtilis* PE7 culture.

Conclusions

PE7

Our study revealed that the plant beneficial bacterium, *B. subtilis* PE7 isolated from kimchi produced IAA and ammonia under *in vitro* conditions, and significantly enhanced tomato growth under *in vivo* conditions. Further studies such as investigation of antimicrobial activity of strain PE7 against phytopathogens, observation of its biocontrol effect on tomato diseases and evaluation of effect of strain PE7 culture on tomato growth under field conditions should be carried out. Based on our results, the application of *B. subtilis* PE7 culture through inoculation

in fertilizer-based medium could be used as a potential eco-friendly approach for plant growth promotion to reduce frequent use of synthetic fertilizer in crop production.

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References

- Ahmed, W., G. Zhou, J. Yang, S. Munir, A. Ahmed, Q. Liu, Z. Zhao, and G. Ji. 2022. *Bacillus amyloliquefaciens* WS-10 as a potential plant growth-promoter and biocontrol agent for bacterial wilt disease of flue-cured tobacco. Egypt. J. Biol. Pest Control 32(1):1-14.
- Alotaibi, F., M.S. Arnaud, and M. Hijri. 2022. In-depth characterization of plant growth promotion potentials of selected alkanes-degrading plant growth-promoting bacterial isolates. Front. Microbiol. 13:863702.
- Bessai, S.A., L. Bensidhoum, and E.H. Nabti. 2022. Optimization of IAA production by telluric bacteria isolated from northern Algeria. Biocatal. Agric. Biotechnol. 41:102319.
- Castillo-Alfonso, F., A. Quintana-Menéndez, G. Vigueras-Ramírez, A.M. Sales-Cruz, L.M. Rosales-Colunga, and R. Olivares-Hernández. 2022. Analysis of the propionate metabolism in Bacillus subtilis during 3-indolacetic production. Microorganisms 10(12):2352.
- Cherif-Silini, H., A. Silini, B. Yahiaoui, I. Ouzari, and A. Boudabous. 2016. Phylogenetic and plant-growth-promoting characteristics of *Bacillus* isolated from the wheat rhizosphere. Ann. Microbiol. 66(3):1087-1097.
- Esitken, A., H.E. Yildiz, S. Ercisli, M.F. Donmez, M. Turan, and A. Gunes. 2010. Effects of plant growth promoting bacteria (PGPB) on yield, growth and nutrient contents of organically grown strawberry. Sci. Hortic. 124(1):62-66.
- FAOSTAT. 2021. Production of tomatoes in 2021. World(total)/Production quantity and area harvested/crops/years. Food and Agriculture Organization Corporate Statistical Database. http://www.fao.org/faostat/en/#data/QCL.
- Glick, B.R. 2012. Plant growth-promoting bacteria: Mechanisms and applications. Scientifica 2012:963401.
- Gohil, R.B., V.H. Raval, R.R. Panchal, and K.N. Rajput. 2022. Plant growth-promoting activity of *Bacillus* sp. PG-8 isolated from fermented panchagavya and its effect on the growth of *Arachis hypogea*. Front. Agron. 4:805454.
- Gusmiaty, M.R.A. and R.Y. Payangan. 2019. Production of IAA (*Indole Acetic Acid*) of the rhizosphere fungus in the Suren community forest stand. IOP Conf. Ser.: Earth Environ. Sci. 343:012058.
- Han, S.E., J.Y. Cho, K.Y. Kim, and C.E.H. Maung. 2023. Role of an antagonistic bacterium, *Bacillus subtilis* PE7, in growth promotion of netted melon (*Cucumis melo* L. var. *reticulatus* Naud). Can. J. Microbiol. e-First https://doi. org/10.1139/cjm-2023-0083.
- Hwang, S.H., C.E.H. Maung, J.S. Noh, W.S. Baek, J.Y. Cho, and K.Y. Kim. 2022. Efficiency and mechanisms of action of pelletized compost loaded with *Bacillus velezensis* CE 100 for controlling tomato Fusarium wilt. Biol. Control 176:105088.
- Jeon, H.L., N.K. Lee, S.J. Yang, W.S. Kim, and H.D. Paik. 2017. Probiotic characterization of *Bacillus subtilis* P223 isolated from kimchi. Food Sci. Biotechnol. 26:1641-1648.
- Karthik, C., N. Elangovan, T.S. Kumar, S. Govindharaju, S. Barathi, M. Oves, and P.I. Arulselvi. 2017. Characterization of multifarious plant growth promoting traits of rhizobacterial strain AR6 under Chromium (VI) stress.

Microbiol. Res. 204:65-71.

- Katsenios, N., V. Andreou, P. Sparangis, N. Djordjevic, M. Giannoglou, S. Chanioti, P. Stergiou, M.Z. Xanthou, I. Kakabouki, D. Vlachakis, S. Djordjevic, G. Katsaros, and A. Efthimiadou. 2021. Evaluation of plant growth promoting bacteria strains on growth, yield and quality of industrial tomato. Microorganisms 9(10):2099.
- Kim, T.Y., S.H. Hwang, J.S. Noh, J.Y. Cho, and C.E.H. Maung. 2022. Antifungal potential of *Bacillus velezensis* CE 100 for the control of different *Colletotrichum* species through isolation of active dipeptide, cyclo-(d-phenylalanyl-d-prolyl). Int. J. Mol. Sci. 23(14):7786.
- Kimura, K. and S. Yokoyama. 2019. Trends in the application of *Bacillus* in fermented foods. Curr. Opin. Biotechnol. 56:36-42.
- Kouam, I.D., J. Mabah, P.G. Ntsoli, L. Tchamani, A. Yaouba, B. Katte, and D. Bitom. 2023. Growth promotion potential of *Bacillus* spp. isolates on two tomato (*Solanum lycopersicum* L.) varieties in the West region of Cameroon. Open Agric. 8(1):20220154.
- Lee, K.H., J.H. Lee, H.W. Choi, and D.B. Lee. 2021. Effect of soaking seeds in the *Bacillus velezensis* GH1-13 solution and foliar spray of chlorella vulgaris on the seed potatoes production. Korean J. Soil Sci. Fert. 54(4):413-422.
- Lee, N.K., S.Y. Kim, S.Y. Choi, and H.D. Paik. 2013. Probiotic *Bacillus subtilis* KU201 having antifungal and antimicrobial properties isolated from kimchi. Food Sci. Biotechnol. 22:1-5.
- Luo, H., Y. Bao, and P. Zhu. 2023. Development of a novel functional yogurt rich in lycopene by *Bacillus subtilis*. Food Chem. 407:135142.
- Minuţ, M., M. Diaconu, M. Roşca, P. Cozma, L. Bulgariu, and M. Gavrilescu. 2022. Screening of Azotobacter, Bacillus and Pseudomonas species as plant growth-promoting bacteria. Processes 11(1):80.
- Nithyapriya, S., S. Lalitha, R.Z. Sayyed, M.S. Reddy, D.J. Dailin, H.A.E. Enshasy, N.L. Suriani, and S. Herlambang. 2021. Production, purification, and characterization of bacillibactin siderophore of *Bacillus subtilis* and its application for improvement in plant growth and oil content in sesame. Sustainability 13(10):5394.
- Olanrewaju, O.S., B.R. Glick, and O.O. Babalola. 2017. Mechanisms of action of plant growth promoting bacteria. World J. Microbiol. Biotechnol. 33:1-16.
- Qiao, J., X. Yu, X. Liang, Y. Liu, R. Borriss, and Y. Liu. 2017. Addition of plant-growth-promoting *Bacillus subtilis* PTS-394 on tomato rhizosphere has no durable impact on composition of root microbiome. BMC Microbiol. 17(1):1-12.
- Quinet, M., T. Angosto, F.J.Y. Lisbona, R.B. Gros, S. Bigot, J.P. Martinez, and S. Lutts. 2019. Tomato fruit development and metabolism. Front. Plant Sci. 10:1554.
- Radhakrishnan, R. and I.J. Lee. 2016. Gibberellins producing *Bacillus methylotrophicus* KE2 supports plant growth and enhances nutritional metabolites and food values of lettuce. Plant Physiol. Biochem. 109:181-189.
- Ratnaningsih, H.R., Z. Noviana, T.K. Dewi, S. Loekito, S. Wiyono, A. Gafur, and S. Antonius. 2023. IAA and ACC deaminase producing-bacteria isolated from the rhizosphere of pineapple plants grown under different abiotic and biotic stresses. Heliyon 9(6):E16306.
- Shin, W., M.A. Siddikee, M.M. Joe, A. Benson, K.Y. Kim, G. Selvakumar, Y.Y. Kang, S.Y. Jeon, S. Samaddar, P. Chatterjee, D. Walitang, M. Chanratana, and T. Sa. 2016. Halotolerant plant growth promoting bacteria mediated salinity stress amelioration in plants. Korean J. Soil Sci. Fert. 49(4):355-367.
- Wagi, S. and A. Ahmed. 2019. Bacillus spp.: Potent microfactories of bacterial IAA. PeerJ 7:e7258.
- Zhou, H., Z.H. Ren, X. Zu, X.Y. Yu, H.J. Zhu, X.J. Li, J. Zhong, and E.M. Liu. 2021. Efficacy of plant growthpromoting bacteria *Bacillus cereus* YN917 for biocontrol of rice blast. Front. Microbiol. 12:684888.