Original Research Article

Assessment of Phenolic Content, Saponin Content, and Antioxidant Activities in Gray, Red, and White Adzuki Bean Germplasm: A Multivariate Analysis

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ABSTRACT Seed color is controlled by several genes and is a key trait in determining the metabolite content and biological activities of legume genotypes. In this study, 296 adzuki bean accessions, including 159 grey, 99 red, and 38 white adzuki beans, were grown in Korea. Variations in total phenolic content (TPC), total saponin content (TSC), DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and ferric reducing antioxidant power (FRAP) were assessed and were reported to be in the ranges of 1.52–8.24 mg GAE/g, 14.36–114.22 mg DE/g, 0.23–12.84 mg AAE/g, 1.05–17.66 mg TE/g, and 0.59–13.14 mg AAE/g, respectively, with a wide variation across adzuki beans. Except for DPPH[•] scavenging activity, the average values declined in the order gray > red > white adzuki beans, each demonstrating a significant variation (p < 0.05). White adzuki beans, which showed low metabolite content and antioxidant activity, were clearly separated from the gray and red genotypes using principal component and hierarchical cluster analyses. Moreover, TPC, TSC, and antioxidant activities were strongly correlated, regardless of seed color. Overall, the diversity of the TPC, TSC, and antioxidant activity in a broad population of adzuki bean genotypes was determined. Furthermore, this study found that seed color variation in adzuki beans had a significant effect on the metabolite content and antioxidant activity. Superior accessions with high levels of TPC, TSC, and antioxidant activity were also discovered and could be used for functional plant breeding and human consumption. The findings of this study may be useful for understanding the relationship between seed coat color and metabolite concentration in adzuki beans, paving the way for molecular-level analyses.

Keywords : adzuki bean, antioxidants, polyphenols, saponins, seed color, Vigna angularis

Adzuki beans (*Vigna angularis* (Willd.) Ohwi & Ohashi) have been grown and consumed for centuries throughout East Asian countries including Korea, Japan, and China (Chu *et al.*, 2021a; Ha *et al.*, 2021; Tayade *et al.*, 2022). Adzuki beans are high in nutrition and bioactive chemicals that improve human health. As a result, they have long been utilized as a versatile component in a variety of Asian cuisines such as paste, soup, porridge, and pastries (Lee *et al.*, 2017a; Yousif *et al.*, 2007). Previous research has shown that adzuki beans are good sources of plant-based fats and proteins, as well as minerals including magnesium, iron, potassium, and zinc, all of which are necessary for human wellness (Chu *et al.*, 2021b; Sangsukiam & Duangmal, 2022; Sahasakul *et al.*, 2022). Furthermore, adzuki beans are high in both soluble and insoluble dietary fibers that may help with weight loss and aid food digestion (Popoola *et al.*, 2023). Adzuki beans are also rich sources of secondary metabolites, the most common of which are saponins and polyphenols, which have a synergistic role in regulating reactive radicals in the

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human body (Liu *et al.*, 2017a, 2017b; Liu & Xu, 2015). Several studies have indicated that adzuki bean phenolic acids, flavonoids, and anthocyanins provide a variety of physiological effects, including antioxidants, anti-inflammatory, anticancer, and antidiabetic properties (Kitano-Okada *et al.*, 2012; Kuriya *et al.*, 2023; Lee *et al.*, 2015a; Sahasakul *et al.*, 2022). In general, consuming adzuki beans gives a wide range of health benefits, which has expanded their use in food industries.

Several environmental and genetic factors, including diseases, genotype variation, temperature, post-harvest treatment, and growth conditions, influence the overall metabolite compositions and biological activity of legumes (Hoeck et al., 2000; Oh et al., 2022). Seed-related traits such as seed size, seed weight, cotyledon color, hilum color, and seed coat color are also additional factors that may influence the chemical makeup of legume seeds (Chu et al., 2021a, 2021b; Kim et al., 2011; Lee et al., 2008, 2017b). Adzuki beans come in a variety of seed colors, including red, black, gray, yellow, and white. Seed color variation in adzuki beans, like in other legumes, is regulated by numerous genes, and it is one of the key seed-related characteristics that determine metabolite levels and biological activities (Chu et al., 2021b; Horiuchi et al., 2015; Redden et al., 2009; Zhao et al., 2022). Furthermore, it is one of the qualities that influence consumer and farmer preferences, as well as genotype selection in breeding (Kaga et al., 2008). As a result, studying the effect of seed color variation on seed metabolite and biological activities in a large population of genetic resources is critical. Furthermore, such studies are required to identify adzuki bean genotypes with high metabolite content that are suited for breeding, conservation, and consumption (Chu et al., 2021a; Li et al., 2022).

Previously, some studies have shown the metabolite variations in adzuki beans of varying seed coat colors. Black and red adzuki beans, for example, are known to contain more phenolic acids, anthocyanins, and flavonoids than other colors (Chu *et al.*, 2021b; Ha *et al.*, 2021; Lee *et al.*, 2015b). White adzuki beans, on the other hand, are thought to be high in nutrient composition but poor in secondary metabolite contents (Li *et al.*, 2022; Nagao *et al.*, 2023). Despite these observations, there has been little research that has looked at the impact of seed coat color variation on a large population of adzuki beans. Evaluating the impact of seed coat color variation using a large population of genetic materials could provide a better understanding of its relationship with seed metabolite levels. Moreover, such studies are desirable in order to uncover superior genotypes that could be used for both consumption and breeding as described before (Kaga *et al.*, 2008; Li *et al.*, 2022). In this study, 296 adzuki beans of various colors, including gray, red, and white were cultivated in Korea, and the diversity of total phenolic content, total saponin content, and antioxidant activities were assessed. Moreover, the effect of seed color variation on each of the parameters was evaluated. This study could serve as a foundation for further research into the relationship between seed color, metabolite concentration, and antioxidant activity in adzuki beans, specifically gray, red, and white genotypes.

MATERIALS AND METHOD

Chemicals and reagents

All of the chemicals and reagents used in this study were of the highest quality and used as obtained. Methanol and ethanol were purchased from Fisher Scientific (Pittsburgh, PA, USA), water from Thomas Scientific (Philadelphia, PA, USA), diosgenin from PhytoLab (Vestenbergsgreuth, Germany), and sulfuric acid from DAEJUNG Chemicals (Siheung, Korea). The other chemicals and reagents, including anhydrous sodium carbonate (Na₂CO₃), vanillin, gallic acid, Folin-Ciocalteu phenol reagent, L-ascorbic acid, sodium carbonate, potassium ferricyanide, trichloroacetic acid, ferric chloride, 1,1-diphenyl-2-picrylhydrazyl radical (DPPH[•]), 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt (ABTS), 6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (Trolox), were obtained from Sigma Aldrich (St. Louis, MO, USA).

Seeds collection, cultivation, and sample preparation

The seeds of 296 adzuki bean genetic materials were obtained from the National Agrobiodiversity Center's gene bank (RDA, Jeonju, Korea). Field cultivation of all adzuki bean varieties was carried out on an experimental field at the Center between June and October 2021. Detailed information regarding cultivation conditions can be found in our recently published paper (Desta *et al.*, 2023). Matured seeds were hand-harvested and classified as gray, red, or white genotypes based on seed coat color. Seed samples from each accession were freeze-dried in an LP500 drier (ilSHINBioBase, Dongducheon, Korea). The dried samples were subsequently pulverized using 3 mm stainless TissueLyser II beads (Quagen, Germantown, MD, USA) and stored at -20°C in sealed plastic bags for further analysis.

Seed sample extraction

Seed sample extraction was carried out in accordance with a previously described protocol, with some modifications. Specifically, 0.2 g of powdered sample was combined in triplicate with 2.5 mL of 80% aqueous methanol. The mixture was sonicated for 45 min in the dark at 25 °C. The mixture was then taken off and centrifuged in a high-speed centrifuge (3134 ×g, 15 min) (Labogene, Daejeon, Korea). The upper supernatant was kept, and the extraction procedure was repeated for the residue. The combined supernatant was then used to determine total saponin content, total phenolic content, 1,1-diphenyl-2-picrylhydrazyl radical (DPPH[•]) scavenging activity, ABTS^{•+} scavenging activity, and ferric-reducing antioxidant power.

Total saponin content (TSC)

The TSC was estimated using a previously known vanillinsulfuric acid assay, with minor modifications (Xu & Chang, 2009). In brief, 25 µL of sample extract was combined with an equal amount of freshly prepared 8% vanillin (w/v in ethanol) and 250 µL of 72% sulfuric acid (v/v in water). The mixture was incubated in a water bath at 60°C for 10 min before cooling in an ice bath for 15 min. The absorbance was then measured at 544 nm on an Eon Microplate Spectrophotometer (Bio-Tek, Winooski, VT, USA). Diosgenin (0.10~2.00 mg/mL) was used as a standard to plot calibration curves ($R^2 > 0.999$). TSC was then calculated as milligrams of diosgenin equivalent per gram of dried seed sample (mg DE/g).

Total phenolic content (TPC)

The TPC was calculated using the Folin-Ciocalteu colorimetric technique (Xu & Chang, 2009). In brief, 100 μ L of sample extract was mixed with 100 μ L of Folin-Ciocalteu reagent, and the mixture was incubated at 25 °C in the dark for 3 min. After adding 100 μ L of Na₂CO₃ solution (2%), the mixture was further incubated in the dark for 30 min. Finally, the absorbance was measured at 750 nm using an Eon Microplate Spectrophotometer (Bio-Tek, Winooski, VT, USA). Gallic acid (0.025~0.500 mg/mL) was used as a standard to plot calibration curves. TPC was estimated as milligrams of gallic acid equivalents (mg GAE/g) per gram of dried material.

Antioxidant activities

Three in vitro experiments were used to evaluate the antioxidant activity of each adzuki bean, as detailed below, using our previously published methodology (Desta *et al.*, 2022). During each experiment, analysis was conducted in triplicate and absorbance was measured with an Eon Microplate Spectro-photometer (Bio-Tek, Winooski, VT, USA).

DPPH[•] scavenging activity

During the DPPH[•] scavenging activity assay, 100 μ L of each sample extract was combined with an equivalent volume of freshly made DPPH-solution (150 μ M). The mixture was then incubated for 30 min in the dark at 25 °C, and the absorbance was measured at 517 nm. Calibration curves were plotted using ascorbic acid (2.5~25.0 mg/L, R² > 0.999), and DPPH[•] scavenging activity was calculated as milligrams of ascorbic acid equivalent per gram of dried material (mg AAE/g).

ABTS*+ scavenging activity

To determine the ABTS^{•+} scavenging activity, 10 μ L of sample extract was combined with 150 μ L of ABTS^{•+} working solution before being incubated for 3 min at 25°C in the dark. Then, the absorbance was measured at 734 nm. Trolox was used as a standard to plot calibration curves (10~250 mg/L, R² > 0.999). ABTS^{•+} scavenging activity was calculated as milligrams of Trolox equivalent per gram of dry material (mg TE/g).

Ferric reducing antioxidant power (FRAP)

For the FRAP test, 60 μ L of sample extract was mixed with 150 μ L of freshly prepared phosphate buffer (pH: 6.6, 0.2 M) and an equal volume of 1% potassium ferricyanide solution (K₃Fe(CN)₆). After 20 min of incubation at 50 °C, 150 μ L of 10% trichloroacetic acid was added. Then, the mixture was centrifuged (3134 × g, 10 min), and 100 μ L of the supernatant was combined with an equal volume of distilled water and 20 μ L of 0.1% ferric chloride solution. After 10 min, the absorbance at 700 nm was measured. Calibration curves were constructed using ascorbic acid (10~150 mg/L, R² > 0.999), and FRAP was presented in mg AAE/g.

Data analysis

Measurements were taken in triplicate during each experiment, and results are reported as mean standard \pm standard deviation

 Table 1. Statistical values on the variations of total phenolic content, total saponin content, and antioxidant activities in gray, red, and white adzuki beans grown in Korea.

Variable	Seed coat color	Number of accessions	Minimum	Maximum	Mean	Variation coefficient (%)	Skewness	Kurtosis
	Gray	159	15.743	108.239	50.937 ^a	41.162	0.694	-0.032
TSC	Red	99	23.258	114.218	42.068 ^b	32.146	2.080	7.447
(mg DE/g)	White	38	14.364	33.850	22.094 ^c	21.325	0.409	-0.369
	Total	296	14.364	114.218	44.268	44.542	1.048	0.988
	Gray	159	2.145	8.235	4.161 ^a	22.262	1.483	3.902
TPC	Red	99	2.702	6.336	3.787 ^b	14.338	1.439	4.535
(mg GAE/g)	White	38	1.522	2.370	1.965 ^c	8.060	-0.459	1.181
	Total	296	1.522	8.235	3.754	27.469	0.513	2.155
	Gray	159	1.127	11.208	3.909 ^a	40.082	1.634	3.948
DPPH	Red	99	1.196	12.840	4.142 ^a	37.358	2.241	10.123
(mg AAE/g)	White	38	0.227	1.164	0.335 ^b	43.004	5.228	29.798
	Total	296	0.227	12.840	3.528	54.039	0.720	2.694
	Gray	159	2.361	16.801	8.511ª	30.450	0.773	0.978
ABTS	Red	99	3.840	17.660	7.806 ^b	26.760	1.263	4.170
(mg TE/g)	White	38	1.054	3.231	1.915 ^c	26.298	0.347	0.004
	Total	296	1.054	17.660	7.428	41.883	0.099	0.522
	Gray	159	1.366	10.841	4.257 ^a	38.269	1.478	3.587
FRAP	Red	99	1.759	13.139	4.141 ^a	38.073	2.244	9.660
(mg AAE/g)	White	38	0.588	2.563	1.298 ^b	26.473	0.705	4.289
	Total	296	0.588	13.139	3.838	46.787	1.158	3.470

AAE, ascorbic acid equivalent; ABTS, ABTS⁺⁺ scavenging activity; DE: Diosgenin equivalent; DPPH, DPPH⁺ scavenging activity; FRAP: Ferric reducing antioxidant power; GAE: Gallic acid equivalent; TE, Trolox equivalent; TPC: Total phenolic content; TSC: Total saponin content.

Different superscript letters in a row represent significantly different means (p < 0.05).

(SD). Statistical analysis was carried out using the xlstat program (Lumivero, CO, USA). To statistically determine the differences between means, an analysis of variance at p < 0.05 was performed. R-software version 4.2 (www.r-project.org) was used for boxplot, principal component (PCA), hierarchical cluster (HCA), and Pearson's correlation analyses.

RESULTS AND DISCUSSION

Variation of TPC and effect of seed color difference

Adzuki beans are well-known for their phenolic content, which plays an important role in health promotion (Wang *et al.*, 2022). The TPC of all 296 adzuki beans was determined using a colorimetric assay, as previously described. Table S1 (Supplemental material) provides the average TPC for each accession, and Table 1 summarizes the statistical data. The TPC was in the range of 1.52~8.24 mg GAE/g with a mean of 3.75 mg GAE/g and a coefficient of variation (CV) of 27.47%. Among the 296 accessions, 52.03% of the adzuki beans had a TPC greater than the total mean value. The highest TPC was found in accession IT-216286, while the lowest was recorded in accession IT-105698. Previously, several studies assessed the level of TPC in various adzuki bean genotypes. For instance, Lee *et al.* (2017a) and Sung *et al.* (2020) independently investigated 209 adzuki bean landraces and 8 adzuki bean cultivars cultivated in Korea and discovered TPC ranges of 1.1~11.7 and 0.52~4.28 mg GAE/g, respectively. In another study, TPC in 150 adzuki beans grown in Korea ranged from 374.5 to 6596.9 μg/g (Kim *et*

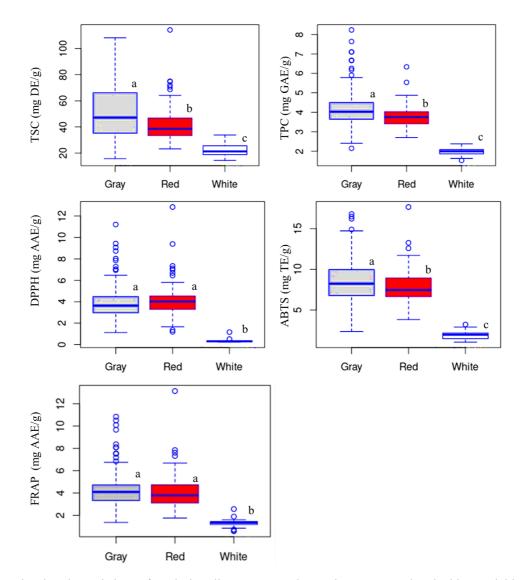


Fig. 1. Boxplots showing the variations of total phenolic content, total saponin content, and antioxidant activities between gray, red, and white adzuki beans. AAE: Ascorbic acid equivalent; ABTS: $ABTS^{*+}$ scavenging activity; DE: Diosgenin equivalent; DPPH: DPPH^{*} scavenging activity; FRAP: Ferric reducing antioxidant power; GAE: Gallic acid equivalent; TE: Trolox equivalent; TPC: Total phenolic content; TSC: Total saponin content. Different letters on boxplots in a category indicate significantly different means (p < 0.05).

al., 2011). Johnson *et al.* (2022) recently discovered a TPC in the range of 53~152 mg GAE/100g in eight adzuki bean genotypes grown in Australia. Several factors, including post-harvest treatment conditions, growing conditions, and analysis techniques, are documented to alter TPC levels in adzuki beans. Therefore, the observed TPC variances in the previous studies could be attributed to these factors (Ha *et al.*, 2021; Szakiel *et al.*, 2011). Overall, the TPC values found in our investigation are within previously reported ranges.

In legumes, seed color has been utilized to discriminate

between metabolite variations. However, there has not been much research on adzuki beans that evaluate the effect of seed color on metabolite levels utilizing a wide population of genotypes (Siah *et al.*, 2014; Wang *et al.*, 2008; Xu *et al.*, 2007). In this study, the TPC variation among the different seed coat colors was statistically evaluated. The box plots in Fig. 1 depict the level of TPC in the adzuki bean accessions according to their seed coat color. Table 1 contains both numerical values and statistical data. TPC ranged from 2.14 to 8.24 mg GAE/g in gray adzuki beans, from 2.70 to 6.34 mg GAE/g in red adzuki beans,

and from 1.52-2.37 mg GAE/g in white adzuki beans. The average TPC decreased in the order of gray (4.16 mg GAE/g) > red (3.79 mg GAE/g) > white 2.37 (mg GAE/g), with significant variation between each group (p < 0.05). Among the accessions with a TPC above the total mean (3.75 mg GAE/g), 67.74% were gray while the remaining 32.26% were red adzuki beans. The TPC of all the white adzuki beans was less than the total mean. Previously, Hori *et al.* (2006) found a much lower TPC in white adzuki beans compared to black, red, and green adzuki beans, which our findings agree with. A recent study also showed white adzuki beans to have a much lower TPC level compared to other colored genotypes (Nagao *et al.*, 2023). In general, our findings along with these previous studies demonstrate that white adzuki beans might not be good sources of polyphenols compared to other colored genotypes.

Variation of TSC and effect of seed color difference

Saponins, found in high concentrations in Adzuki beans, work in tandem with polyphenols to prevent disease and improve health (Liu et al., 2017b). In this study, the TSC of all accessions was determined using the vanillin-H₂SO₄ assay as described before. With a mean of 44.27 mg DE/g, the TSC in the entire population was in the range of 14.36~114.22 mg DE/g, indicating an approximately 8-fold variation across the adzuki bean accessions (CV = 44.27%) (Table S1, Table 1). Around 40.88% of the adzuki bean accessions had a TSC greater than the total mean. Accession IT-194575 had the highest TSC, while accession IT-294630 had the lowest. Many studies have shown that saponin concentrations differ amongst adzuki bean genotypes, and genetic variations, post-harvest management, extraction procedures, and growing conditions can all impact TSC levels. Previously, TSC levels in two adzuki bean genotypes ranged from 10.33 to 19.40 mg/g after different sonication treatments (Chiu, 2021). In other studies, Luo et al. (2016) and Liu & Xu (2015) separately reported TSC of 45.60 mg DE/g and 10.82 mg SBaE/g in two different Chinese adzuki beans. The TSC value in the former study corresponded to the total mean value in our investigation.

Saponin levels in legumes are also affected by seed color variations. However only a few studies were published in adzuki beans (Siah *et al.*, 2014; Wang *et al.*, 2008; Xu *et al.*, 2007). In this study, the effect of seed color variation on TSC was statistically examined, and significant variation was found (Fig.

1, Table 1). TSC in gray, red, and white adzuki beans ranged from 15.74 to 108.24, 23.46 to 114.22, and 14.36 to 33.85 mg DE/g, respectively. The average TSC level followed a similar pattern as TPC, being the highest in gray adzuki beans (50.94 mg DE/g) and the lowest in white adzuki beans (22.09 mg DE/g). Statistically, the average TSC varied significantly among the different seed coat colors (p < 0.05). Moreover, gray adzuki beans accounted for 72.73% of the accessions with a TSC greater than the total mean, whereas red adzuki beans accounted for the remaining 27.27%. Again, none of the white adzuki beans had a TSC greater than the total mean value. To the best of our knowledge, there are no previous studies that investigated the influence of seed coat color on total saponin content in adzuki beans. Overall, our research found that white adzuki beans are low in saponin content when compared to gray and red adzuki beans.

Variations of antioxidant activities and effect of seed color difference

In vitro tests are widely accepted as the most convenient and cost-effective assays for determining the total antioxidant activity of plant and food extracts. However, due to variances in the scavenging mechanism, a single experiment alone cannot be utilized to evaluate the total antioxidant activity of such extracts (Martins et al., 2022). In this study, three separate assays were utilized to evaluate the antioxidant activity of each adzuki bean accession (Table 1, Table S1). The DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and FRAP were in the ranges of 0.23~12.84 mg AAE/g, 1.05~17.66 mg TE/g, and 0.59~13.14 mg AAE/g, respectively signifying a wide range of variations among the adzuki bean accessions. Accession IT-194575 displayed the highest DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and FRAP at the same time. This accession had the highest TSC as well as a TPC of 6.34 mg GAE/g, which is the 7th highest level of all the 296 accessions. On the other hand, accessions IT-236259, IT-105890, and IT-196872 displayed the lowest DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and FRAP, respectively. Compared to our study, much lower ABTS^{•+} scavenging activity (1.64~8.15 mg TE/g) and FRAP (0.41~5.44 mg AAE/g) ranges were reported, and the variations could be due to the difference in the number of genotypes investigated (Lee et al., 2017a; Sung et al., 2020). In addition to the studies mentioned above, other researchers have

also revealed the antioxidant activities of a variety of adzuki bean genotypes. However, differences in extraction techniques, reporting methodologies, and tests used made comparing published values to our findings difficult (Gohara *et al.*, 2016; Han *et al.*, 2022). In general, our findings demonstrated wide variations of antioxidant activities among the different adzuki beans. Furthermore, accessions with greater levels of TPC and TSC displayed stronger antioxidant activities than those with lower levels of TPC and TSC (Table S1). Such findings have been reported in numerous prior research, indicating the roles of polyphenols and saponins in the regulation of reactive radicals (Johnson *et al.*, 2022; Lee *et al.*, 2017a).

The effect of seed coat color variations on each of the antioxidant activities was also statistically evaluated (Table 1, Fig. 1). The DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and FRAP were in the ranges of 1.13~11.21 mg AAE/g, 2.36~ 16.80 mg TE/g, and 1.37~10.84 mg AAE/g in gray, 1.20~12.84 mg AAE/g, 3.84~17.66 mg TE/g, and 1.76~13.14 mg AAE/g in red, and 0.23~1.64 mg AAE/g, 1.05~3.23 mg TE/g, and 0.59~2.56 mg AAE/g in white adzuki beans, respectively. The average ABTS⁺⁺ scavenging activity and FRAP each decreased in the order of gray > red > white, while DPPH[•] scavenging activity decreased in the order of red (4.14 mg AAE/g) > gray (3.91 mg AAE/g) > white (0.34 mg AAE/g) adzuki beans (Table 1, Fig. 1). ABTS⁺⁺ scavenging activity differed significantly between the three seed coat colors. Likewise, DPPH[•] scavenging activity and FRAP of gray and red adzuki beans differed significantly from those of white adzuki beans but not from each other. Furthermore, gray adzuki beans accounted for 56.38, 72.85, and 65.96% of the accessions with values above the total means of DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and FRAP, respectively, whereas red adzuki beans accounted for the remaining 43.62, 27.15, and 34.04%, respectively. For any of the antioxidant activities, no white adzuki bean exceeded the total mean values. Little research has been done on the effects of seed color on antioxidant activity in adzuki beans. In line with our findings, a recent investigation showed much lower DPPH[•] scavenging activity in white adzuki beans as compared to other colored genotypes, including black and red adzuki beans (Nagao et al., 2023). In another study, Kim et al. (2015) observed that black adzuki beans had higher DPPH[•] scavenging activity and ABTS*+ scavenging activity than red adzuki beans, further signifying the influence of seed color. In general, our findings demonstrated that seed coat color variation had a significant effect on adzuki bean antioxidant activities. As a result, it can be concluded that white adzuki beans do not have the same antioxidant content as gray and red adzuki beans.

Multivariate analysis

To gain a better understanding of the changes in total metabolite contents and antioxidant activity among the adzuki beans studied, hierarchical cluster analysis (HCA), principal component analysis (PCA), and Pearson's correlation analysis were conducted. The HCA in Fig. 2A clearly shows the low levels of TPC, TSC, and antioxidant activities in white adzuki beans. This result was reinforced further by the PCA, which was computed along the first two principal components (PC1 and PC2), which together contributed more than 90% of the total variance. As shown in the score plot, white adzuki beans were differentiated from gray and red adzuki beans (Fig. 2B). The gray and red landraces, on the other hand, were widely scattered with one another. As indicated in Fig. 2C and Table 2, TPC (20.44%), DPPH[•] scavenging activity (20.67%), ABTS^{•+} scavenging activity (21.85%), and FRAP (21.03%) contributed the most to the variations observed along PC1, while TSC (74.12%) contributed the most along PC2. This was consistent with the HCA, which showed that TPC, ABTS⁺⁺ scavenging activity, DPPH[•] scavenging activity, and FRAP were all clustered together (Fig. 2A). In general, the PCA supported by HCA in our study clearly separated white adzuki beans from gray and red accessions. As a result, TSC, TPC, ABTS^{•+} scavenging activity, DPPH[•] scavenging activity, and FRAP could be important traits for distinguishing a diverse population of adzuki bean genetic materials with varying seed coat colors.

Pearson's correlation analysis was performed to compute the correlation between the analyzed parameters. Positive and strong correlations were found between the examined variables in all the adzuki beans. However, the correlation coefficient (r) values were greater in gray (Fig. 3A) and red (Fig. 3B) adzuki beans than in white (Fig. 3C) adzuki beans. In gray adzuki beans, for example, TPC had r = 0.81 with DPPH[•] scavenging activity, r = 0.76 with ABTS^{•+} scavenging activity, and r = 0.63 with FRAP, all of which were significant at p < 0.001. However, in white adzuki beans, the correlation of TPC with DPPH[•] scavenging activity, and FRAP was considerably weaker, with r of 0.58, 0.26, and 0.48,

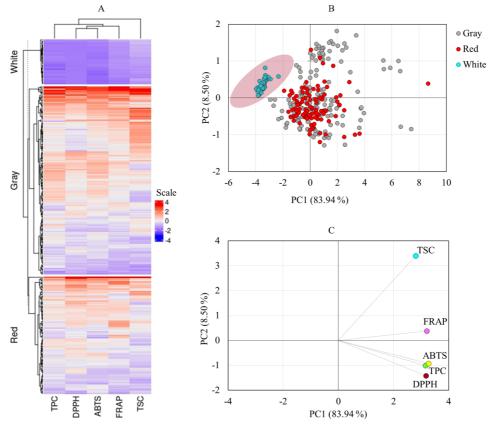


Fig. 2. Hierarchical cluster analysis (A), score plot of adzuki beans (B), and loading plot of variables (C) from principal component analysis. ABTS, ABTS⁺⁺ scavenging activity; DPPH, DPPH⁺ scavenging activity; FRAP: Ferric reducing antioxidant power; TPC: Total phenolic content; TSC: Total saponin content.

Table 2. Total phenolic content, total saponin content, and antioxidant activity of top-performing adzuki bean accessions.

TSC (mg DE/g)	TPC (mg GAE/g)	DPPH (mg AAE/g)	ABTS (mg TE/g)	FRAP (mg AAE/g)
IT No. Color Content	IT No. Color Content	IT No. Color Activity	IT No. Color Activity	IT No. Color Activity
194575 Red 114.22	216286 Gray 8.24	194575 Red 12.84	194575 Red 17.66	194575 Red 13.14
216285 Gray 108.24	216306 Gray 7.64	216286 Gray 11.21	216306 Gray 16.80	216285 Gray 10.84
216305 Gray 107.21	216305 Gray 7.11	216285 Gray 9.43	216305 Gray 16.48	216286 Gray 10.50
216303 Gray 106.44	216303 Gray 7.10	103973 Red 9.39	216285 Gray 16.21	216306 Gray 10.09
216298 Gray 106.16	216299 Gray 6.68	103060 Gray 9.06	216298 Gray 14.90	216305 Gray 9.67
216299 Gray 100.91	216298 Gray 6.66	216306 Gray 8.74	216299 Gray 14.73	216298 Gray 8.37
240343 Gray 100.55	194575 Red 6.34	216298 Gray 8.03	216286 Gray 14.54	196895 Gray 8.10
211854 Gray 91.36	178391 Gray 6.26	216305 Gray 7.86	178391 Gray 14.27	216299 Gray 8.09
283300 Gray 90.28	216300 Gray 6.18	105643 Red 7.34	216303 Gray 14.21	178458 Red 7.85
208913 Gray 89.64	216289 Gray 5.91	216291 Gray 7.28	216300 Gray 14.12	178527 Red 7.61
216286 Gray 87.61	216291 Gray 5.79	104171 Red 7.12	186309 Red 13.27	216291 Gray 7.55
252929 Gray 85.38	196895 Gray 5.55	104855 Gray 7.04	196895 Gray 13.17	216303 Gray 7.51
280309 Gray 84.16	186309 Red 5.54	186309 Red 6.98	216295 Gray 12.82	196870 Red 7.32
195537 Gray 82.80	186322 Gray 5.52	216299 Gray 6.94	103973 Red 12.59	216300 Gray 7.18
208790 Gray 82.46	121459 Gray 5.37	105632 Red 6.69	186322 Gray 12.54	216295 Gray 6.85

AAE, ascorbic acid equivalent; ABTS, ABTS⁺⁺ scavenging activity; DE: Diosgenin equivalent; DPPH, DPPH⁺ scavenging activity; FRAP: Ferric reducing antioxidant power; GAE: Gallic acid equivalent; TE, Trolox equivalent; TPC: Total phenolic content; TSC: Total saponin content.

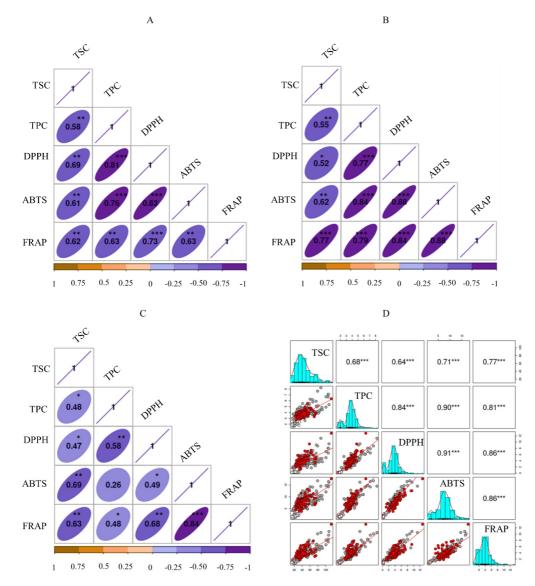


Fig. 3. Pearson correlation matrix of metabolite contents and antioxidant activities in gray (A), red (B), and white (C) adzuki beans, and the entire dataset (D). ABTS, $ABTS^{*+}$ scavenging activity; DPPH, DPPH[•] scavenging activity; FRAP: Ferric reducing antioxidant power; TPC: Total phenolic content; TSC: Total saponin content. *p < 0.05, **p < 0.01, ***p < 0.001.

respectively. TSC also correlated with antioxidant activities in a similar way as TPC, with $r \ge 0.61$ in gray, $r \ge 0.52$ in red, and $r \ge 0.47$ in white adzuki beans. Generally, TSC and TPC demonstrated positive associations with all antioxidant activities and with each other regardless of seed coat color (Fig. 3D), with all except the TPC link with ABTS^{•+} scavenging activity in white adzuki beans being significant. The observed strong and positive correlations were consistent with several prior research (Han *et al.*, 2022; Lee *et al.*, 2017a).

To summarize, this study investigated the variations of total phenolic content, total saponin content and antioxidant activities among a large population of adzuki bean accessions cultivated in Korea. All the variables studied including total phenolic content, total saponin content, DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and ferric-reducing antioxidant power demonstrated a large range of variability among the adzuki beans. Furthermore, seed coat color variation had a significant effect on all of the parameters, with gray adzuki beans having the highest average values of all except DPPH[•] scavenging activity and white landraces having the lowest levels. As previously indicated, adzuki beans with high levels of TPC and TSC are useful due to their multiple health benefits (Li *et al.*, 2022). This

Variables	PC1	PC2	PC3	PC4	PC5
TSC	16.020	74.117	2.062	6.558	1.243
TPC	20.438	6.557	47.661	14.271	11.073
DPPH	20.666	12.874	15.995	25.046	25.419
ABTS	21.848	5.525	2.335	9.776	60.516
FRAP	21.027	0.928	31.948	44.348	1.749
Eigenvalue	4.197	0.425	0.188	0.118	0.072
Variability (%)	83.936	8.505	3.759	2.358	1.443
Cumulative (%)	83.936	92.440	96.199	98.557	100.000

Table 3. Eigenvalues and individual and cumulative contributions of variables in the first five principal components.

ABTS, ABTS⁺⁺ scavenging activity; DPPH, DPPH[•] scavenging activity; FRAP: Ferric reducing antioxidant power; TPC: Total phenolic content; TSC: Total saponin content. PC1: First principal component; PC2: Second principal component; PC3: Third principal component; PC4: Fourth principal component; PC5: Fifth principal component.

study also identified the top 15 superior adzuki beans showing the highest levels of each parameter (Table 3). In particular, accessions IT-194575, IT-216286, IT-216298, IT-216299, and IT-216305 contained high levels of total phenolic content, total saponin content, DPPH[•] scavenging activity, ABTS^{•+} scavenging activity, and ferric-reducing antioxidant power all at the same time. Accession IT-216303, on the other hand, showed high levels of all parameters except DPPH[•] scavenging activity, whereas accession IT-216306 showed high levels of all parameters except total saponin content. Accession IT-216300 likewise had high levels of all parameters except total saponin content and DPPH[•] scavenging activity. As a result, these accessions have the potential to be valuable sources of healthpromoting metabolites. Furthermore, the findings of this study may inspire further molecular research in the future. Identifying specific saponins and polyphenol components in gray, red, and white adzuki beans, on the other hand, would also be a good future research endeavor.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- Chiu, K. Y. 2021. Changes in microstructure, germination, sprout growth, phytochemical and microbial quality of ultrasonication treated adzuki bean seeds. Agronomy 11(6) : 1093. https://doi.org/10.3390/agronomy11061093
- Chu, L., P. Zhao, X. Huang, B. Zhao, Y. Li, K. Yang, and P. Wan. 2021a. Genetic analysis of seed coat colour in adzuki bean (*Vigna angularis* L.). Plant Genetic Resources : Characterisation and Utilisation 19(1) : 67-73. https://doi.org/10.1017/S1479 262121000101
- Chu, L., P. Zhao, K. Wang, B. Zhao, Y. Li, K. Yang, and P. Wan. 2021b. VaSDC1 Is Involved in Modulation of Flavonoid Metabolic Pathways in Black and Red Seed Coats in Adzuki Bean (*Vigna angularis* L.). Frontiers in Plant Science 12 : 679892. https://doi.org/10.3389/fpls.2021.679892
- Desta, K. T., O. S. Hur, S. Lee, H. Yoon, M.-J. Shin, J. Yi, Y. Lee, N. Y. Ro, X. Wang, and Y.-M. Choi. 2022. Origin and seed coat color differently affect the concentrations of metabolites and antioxidant activities in soybean (*Glycine max* (L.) Merrill) seeds. Food Chemistry 381 : 132249. https://doi.org/ 10.1016/j.foodchem.2022.132249
- Desta, K. T., Y.-M. Choi, J. Yi, S. Lee, M.-J Shin, and X. Wang. 2023. Agro-morphological Characterization of Korean, Chinese, and Japanese Adzuki Bean (*Vigna angularis* (Willd.) Ohwi & Ohashi) Genotypes. Korean Journal of Crop Science 68(1): 8-19. https://doi.org/10.7740/kjcs.2023.68.1.008
- Gohara, A. K., A. H. Souza, S. T. Gomes, N. E. Souza, J. V. Visentainer, and M. Matsushita. 2016. Nutritional and bioactive compounds of adzuki beans cultivars using chemometric approach. Ciencia E. Agrotecnologia 40(1): 104-113. http://dx.doi.org/10.1590/S1413-70542016000100010
- Ha, T. J., J. E. Park, K. S. Lee, W. D. Seo, S. B. Song, M. H. Lee, S. Kim, J. I. Kim, E. Oh, S. B. Pae, D. Y. Kwak, and J. H. Lee.

2021. Identification of anthocyanin compositions in black seed coated Korean adzuki bean (*Vigna angularis*) by NMR and UPLC-Q-Orbitrap-MS/MS and screening for their antioxidant properties using different solvent systems. Food Chemistry 346 : 128882. https://doi.org/10.1016/j.foodchem. 2020.128882

- Han, N., K. S. Woo, J. Y. Lee, S. B. Song, Y. Y. Lee, M. Kim, M. S. Kang, and H. J. Kim. 2022. Comparison of Physicochemical Characteristics, Functional Compounds, and Physiological Activities in Adzuki Bean Cultivars. Journal of the Korean Society of Food Science and Nutrition 51(5): 428-438. https:// doi.org/10.3746/jkfn.2022.51.5.428
- Hoeck, J. A., W. R. Fehr, P. A. Murphy, and G. A. Welke. 2000. Influence of genotype and environment on isoflavone contents of soybean. Crop Science 40(1): 48-51. https://doi.org/10.2135/ cropsci2000.40148x
- Hori, Y., S. Sato, and A. Hatai. 2006. Antibacterial activity of plant extracts from azuki beans (*Vigna angularis*) in vitro. Phytotherapy Research 20(2): 162-164. https://doi.org/10.1002/ ptr.1826
- Horiuchi, Y., H. Yamamoto, R. Ogura, N. Shimoda, H. Sato, and K. Kato. 2015. Genetic Analysis and Molecular Mapping of Genes Controlling Seed Coat Colour in Adzuki Bean (*Vigna angularis*). Euphytica 206(3): 609-617. https://doi.org/10.1007/ s10681-015-1461-9.
- Johnson, J. B., P. Neupane, S. P. Bhattarai, T. Trotter, and M. Naiker. 2022. Partitioning of nutritional and phytochemical constituents in nine Adzuki bean genotypes from Australia. Journal of Agriculture and Food Research 10 : 100398. https://doi.org/10.1016/j.jafr.2022.100398
- Kaga, A., T. Isemura, N. Tomooka, and D. A. Vaughan. 2008. The genetics of domestication of the azuki bean (*Vigna angularis*). Genetics 178(2): 1013-1036. https://doi.org/10.1534/genetics. 107.078451
- Kim, E.-H., H.-K. Song, Y.-J. Park, J.-R. Lee, M.-Y. Kim, and I.-M. Chung. 2011. Determination of Phenolic Compounds in Adzuki bean (*Vigna angularis*) Germplasm. Korean Journal of Crop Science 56(4): 375-384. https://doi.org/10.7740/kjcs. 2011.56.4.375
- Kim, M., J.-E. Park, S.-B. Song, and Y.-S. Cha. 2015. Effect of black adzuki bean (*Vigna angularis*) extract on proliferation and differentiation of 3T3-L1 preadipocytes into mature adipocytes. Nutrients 7 : 277-292. https://doi.org/10.3390/nu 7010277
- Kitano-Okada, T., A. Ito, A. Koide, Y. Nakamura, K. H. Han, K. Shimada, K. Sasaki, K. Ohba, S. Sibayama, and M. Fukushima. 2012. Anti-obesity role of adzuki bean extract containing polyphenols: In vivo and in vitro effects. Journal of the Science of Food and Agriculture 92(13) : 2644-2651. https://doi.org/10. 1002/jsfa.5680
- Kuriya, K., S. Goto, E. Kobayashi, M. Nishio, M. Nakamura, and H. Umekawa. 2023. Cholesterol-lowering activity of adzuki

bean (*Vigna angularis*) polyphenols. Molecular Biology Reports 50(7): 5575-5584. https://doi.org/10.1007/s11033-023-08481-7

- Lee, K. J., K. H. Ma, Y. H. Cho, J. R. Lee, J. W. Chung, and G. A. Lee. 2017a. Phytochemical distribution and antioxidant activities of Korean adzuki bean (*Vigna angularis*) landraces. Journal of Crop Science and Biotechnology 20(3) : 205-212. https://doi.org/10.1007/s12892-017-0056-0
- Lee, J., Y. S. Hwang, S. T. Kim, W. B. Yoon, W. Y. Han, I. K. Kang, and M. G. Choung. 2017b. Seed coat color and seed weight contribute differential responses of targeted metabolites in soybean seeds. Food Chemistry 214 : 248-258. https://doi. org/10.1016/j.foodchem.2016.07.066
- Lee, L. S., E. J. Choi, C. H. Kim, J. M. Sung, Y. B. Kim, J. S. Kum, and J. D. Park. 2015b. Antioxidant properties of different parts of red and black adzuki beans. Journal of the Korean Society of Food Science and Nutrition 44(8) : 1150-1156. https://doi. org/10.3746/jkfn.2015.44.8.1150
- Lee, S. J., J. J. Kim, H. I. Moon, J. K. Ahn, S. C. Chun, W. S. Jung, O. K. Lee, and I. M. Chung. 2008. Analysis of isoflavones and phenolic compounds in Korean soybean [*Glycine max* (L.) Merrill] seeds of different seed weights. Journal of Agricultural and Food Chemistry 56(8): 2751-2758. https://doi.org/10.1021/ jf073153f
- Lee, S. S., N. Mohd Esa, and S. P. Loh. 2015a. In vitro inhibitory activity of selected legumes against pancreatic lipase. Journal of Food Biochemistry 39(4): 485-490. https://doi.org/10.1111/ jfbc.12150
- Li, H, L. Zou, X. Li, D. Wu, H. Liu, H. Li, and R. Gan. 2022. Adzuki bean (*Vigna angularis*): chemical compositions, physicochemical properties, health benefits, and food applications. Comprehensive Review in Food Science and Food Safety 21: 2335-62. https://doi.org/10.1111/1541-4337. 12945
- Liu, R., and B. Xu. 2015. Inhibitory effects of phenolics and saponins from commonly consumed food legumes in China against digestive enzymes pancreatic lipase and α -Glycosidase. International Journal of Food Properties 18(10) : 2246-2255. https://doi.org/10.1080/10942912.2014.971178
- Liu, R., Y. Zheng, Z. Cai, and B. Xu. 2017a. Saponins and flavonoids from adzuki bean (*Vigna angularis* L.) ameliorate high-fat diet-induced obesity in ICR mice. Frontiers in Pharmacology 8 : 1-8. https://doi.org/10.3389/fphar.2017.00687
- Liu, R., Z. Cai, and B. Xu. 2017b. Characterization and quantification of flavonoids and saponins in adzuki bean (*Vigna angularis* L.) by HPLC-DAD-ESI-MSn analysis. Chemistry Central Journal 11(1): 1-17. https://doi.org/10.1186/s13065-017-0317-x
- Luo, J., W. Cai, T. Wu, and B. Xu. 2016. Phytochemical distribution in hull and cotyledon of adzuki bean (*Vigna angularis* L.) and mung bean (*Vigna radiate* L.), and their contribution to antioxidant, anti-inflammatory and anti-diabetic activities. Food Chemistry 201 : 350-360. https://doi.org/10.1016/j.food

chem.2016.01.101

- Martins, G. R., M. M. G. Mattos, F. M. Nascimento, F. L. Brum, R. Mohana-Borges, N. G. Figueiredo, D. F. M. Neto, G. B. Domont, F. C. S. Nogueira, F. D. A. De Paiva Campos, and A. Sant'Ana Da Silva. 2022. Phenolic Profile and Antioxidant Properties in Extracts of Developing Açaí (*Euterpe oleracea* Mart.) Seeds. Journal of Agricultural and Food Chemistry 70(51): 16218-16228. https://doi.org/10.1021/acs.jafc.2c07028
- Nagao, N., Y. Sakuma, T. Funakoshi, and T. Itani. 2023. Variation in antioxidant capacity of the seven azuki bean (*Vigna angularis*) varieties with different seed coat color. Plant Production Science 26(2): 164-173. https://doi.org/10.1080/ 1343943X.2023.2206576
- Oh, S.-M., J. Jang, K. Park, Y. Kang, J.-S. Lee, S.-B. Song, T. Yun, and J. Y. Kim. 2022. Yield and Antioxidant Properties of Korean Adzuki Bean (*Vigna angularis* L.) Cultivars Under Different Air Temperatures and Sunshine Hours. Korean Journal of Crop Science 67(3): 189-197. https://doi.org/10.7740 /kjcs.2022.67.3.189
- Popoola, J. O., O. B. Ojuederie, O. S. Aworunse, A. Adelekan, A. S. Oyelakin, O. L. Oyesola, P. A. Akinduti, S. O. Dahunsi, T. T. Adegboyega, S. U. Oranusi, M. S. Ayilara, and C. A. Omonhinmin. 2023. Nutritional, functional, and bioactive properties of African underutilized legumes. Frontiers in Plant Science 14: 1105364. https://doi.org/10.3389/fpls.2023.1105364
- Redden, R. J., K. E. Basford, P. M. Kroonenberg, F. M. A. Islam, R. Ellis, S. Wang, Y. Cao, X. Zong, and X. Wang. 2009. Variation in Adzuki Bean (*Vigna angularis*) Germplasm Grown in China. Crop Science 49(3) : 771-782. https://doi. org/10.2135/cropsci2008.03.0175.
- Sahasakul, Y., A. Aursalung, S.Thangsiri, P. Wongchang, P. Sangkasa-ad, A. Wongpia, A. Polpanit, W. Inthachat, P. Temviriyanukul, and U. Suttisansanee. 2022. Nutritional Compositions, Phenolic Contents, and Antioxidant Potentials of Ten Original Lineage Beans in Thailand. Foods 11(14) : 2062. https://doi.org/10.3390/foods11142062
- Sangsukiam, T., and K. Duangmal. 2022. Changes in bioactive compounds and health-promoting activities in adzuki bean: Effect of cooking conditions and in vitro simulated gastrointestinal digestion. Food Research International 157 : 111371. https://doi.org/10.1016/j.foodres.2022.111371
- Siah, S., J. A. Wood, S. Agboola, I. Konczak, and C. L. Blanchard. 2014. Effects of soaking, boiling and autoclaving on the

phenolic contents and antioxidant activities of faba beans (*Vicia faba* 1.) differing in seed coat colours. Food Chemistry 142:461-468. https://doi.org/10.1016/j.foodchem.2013.07.068

- Sung, J. S., S. B. Song, J. Y. Kim, Y. J. An, J. E. Park, M. E. Choe, and J. H. Chu. 2020. Variation in Physicochemical Characteristics and Antioxidant Activities of Small Redbean Cultivars. Korean Journal of Crop Science 65(3) : 231-240. https://doi. org/10.7740/kjcs.2020.65.3.231
- Szakiel, A., C. Pączkowski, and M. Henry. 2011. Influence of environmental abiotic factors on the content of saponins in plants. Phytochemistry Reviews 10(4) : 471-491. https://doi. org/10.1007/s11101-010-9177-x
- Tayade, R., S. Kim, P. Tripathi, Y. Choi, J. Yoon, and Y. Kim. 2022. High-throughput Root Imaging Analysis Reveals Wide. Plants 11 : 405. https://doi.org/10.3390/plants11030405
- Wang, M. L., A. G. Gillaspie, J. B. Morris, R. N. Pittman, J. Davis, and G. A. Pederson. 2008. Flavonoid content in different legume germplasm seeds quantified by HPLC. Plant Genetic Resources : Characterisation and Utilisation 6(1) : 62-69. https:// doi.org/10.1017/S1479262108923807
- Wang, Y., X. Yao, H. Shen, R. Zhao, Z. Li, X. Shen, F. Wang, K. Chen, Y. Zhou, B. Li, X. Zheng, and S. Lu. 2022. Nutritional Composition, Efficacy, and Processing of *Vigna angularis* (Adzuki Bean) for the Human Diet: An Overview. Molecules 27(18) : 6079. https://doi.org/10.3390/molecules27186079
- Xu, B. J., S. H. Yuan, and S. K. C. Chang. 2007. Comparative analyses of phenolic composition, antioxidant capacity, and color of cool season legumes and other selected food legumes. Journal of Food Science 72(2) : S167-S177. https://doi.org/10. 1111/j.1750-3841.2006.00261.x
- Xu, B., and S. K. C. Chang. 2009. Phytochemical profiles and health-promoting effects of cool-season food legumes as influenced by thermal processing. Journal of Agricultural and Food Chemistry 57(22): 10718-10731. https://doi.org/10.1021/ jf902594m
- Yousif, A. M., J. Kato, and H. C. Deeth. 2007. Effect of Storage on the Biochemical Structure and Processing Quality of Adzuki Bean (*Vigna angularis*). Food Rev. Int. 23 : 1-33
- Zhao, P., L. Chu, K. Wang, B. Zhao, Y. Li, K. Yang, and P. Wan. 2022. Analyses on the pigment composition of different seed coat colors in adzuki bean. Food Science and Nutrition 10(8): 2611-2619. https://doi.org/10.1002/fsn3.2866

Supplementary Table 1. General information and average total metabolite content and antioxidant activity of 296 Adzuki bean accessions grown in Korea.

	Ger	neral information	1		Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg_GAE/g)	TSC (mg_DE/g)	DPPH (mg_AAE/g)	ABTS (mg_TE/g)	FRAP (mg AAE/g)
25924	JPN	Unknown	Red	4.00	41.68	4.23	7.96	4.32
100899	KOR	Landrace	Red	4.00	34.84	3.46	7.96	3.30
100916	KOR	Landrace	Red	3.84	35.37	3.06	7.24	3.60
100961	KOR	Landrace	Gray	2.76	23.50	2.52	4.48	1.78
100997	KOR	Landrace	Red	4.27	45.35	4.56	9.00	4.56
101052	KOR	Landrace	Red	4.03	42.15	4.10	8.95	3.72
101083	KOR	Landrace	Red	4.04	40.75	4.09	8.93	4.14
101098	KOR	Landrace	Red	3.38	23.26	2.70	6.14	2.95
101355	KOR	Landrace	Gray	3.11	26.69	3.53	6.26	2.58
102814	KOR	Landrace	Red	3.72	46.72	4.09	8.25	4.16
102856	KOR	Landrace	Red	4.59	48.50	5.23	10.83	5.82
102970	KOR	Landrace	Red	3.79	30.25	2.37	5.37	2.88
102977	KOR	Landrace	Red	3.50	43.79	3.07	7.33	3.91
103037	KOR	Landrace	Red	3.56	24.41	1.95	4.64	2.59
103054	KOR	Landrace	Red	3.94	45.13	4.02	8.17	4.42
103060	KOR	Landrace	Gray	5.08	76.34	9.06	12.53	5.58
103329	KOR	Landrace	Red	3.63	34.39	2.37	5.64	3.11
103372	KOR	Landrace	Red	4.02	37.01	3.75	7.76	4.20
103420	KOR	Landrace	Gray	3.37	33.36	4.28	6.64	2.81
103427	KOR	Landrace	Gray	3.63	42.70	4.50	7.63	3.01
103581	KOR	Landrace	Gray	3.48	38.12	4.27	7.50	2.89
103602	KOR	Landrace	Gray	2.82	33.62	3.29	6.49	2.33
103603	KOR	Landrace	Red	3.45	34.57	2.25	5.49	2.79
103719	KOR	Landrace	Red	4.32	40.23	3.46	7.29	3.74
103751	KOR	Landrace	Red	3.83	47.04	3.02	6.74	3.54
103771	KOR	Landrace	Red	3.62	37.98	2.55	5.22	2.80
103937	KOR	Landrace	Red	3.84	28.68	5.72	9.17	4.69
103953	KOR	Landrace	Red	4.07	30.22	4.42	7.44	4.15
103962	KOR	Landrace	Red	3.95	32.93	5.39	8.86	4.69
103972	KOR	Landrace	Red	3.95	25.62	4.07	7.95	4.03
103973	KOR	Landrace	Red	4.77	58.52	9.39	12.59	6.69
104019	KOR	Landrace	Red	3.91	44.49	5.08	9.51	4.77
104158	KOR	Landrace	Red	3.81	33.17	3.38	6.80	3.60
104171	KOR	Landrace	Red	4.88	46.94	7.12	11.71	5.64
104203	KOR	Landrace	White	1.75	20.74	0.34	1.57	1.37
104275	KOR	Landrace	Gray	2.40	20.46	1.67	3.80	1.37
104301	KOR	Landrace	White	1.80	31.54	0.26	2.08	1.39
104309	KOR	Landrace	Gray	3.32	32.44	2.54	5.17	1.93
104400	KOR	Landrace	Gray	3.71	26.80	2.47	5.43	1.86
104439	KOR	Landrace	Red	3.35	31.93	2.93	7.65	2.60

	Gei	neral informatior	1		Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg GAE/g)	TSC (mg DE/g)	DPPH (mg AAE/g)	ABTS (mg TE/g)	FRAP (mg AAE/g
104459	KOR	Landrace	Red	3.37	34.57	3.89	9.08	3.07
104530	KOR	Landrace	Red	3.85	37.91	4.07	8.93	3.15
104547	KOR	Landrace	Red	3.58	36.38	3.90	8.06	2.63
104549	KOR	Landrace	Red	4.28	52.02	5.47	9.81	3.63
104552	KOR	Landrace	Red	4.15	47.13	4.58	9.84	3.70
104753	KOR	Landrace	Red	3.42	38.97	4.05	5.34	2.50
104814	KOR	Landrace	Red	3.42	31.61	3.96	5.29	2.39
104825	KOR	Landrace	Red	3.28	33.52	4.03	5.14	2.40
104833	KOR	Landrace	Red	3.40	37.72	3.81	5.15	2.43
104855	KOR	Landrace	Gray	4.63	54.72	7.04	12.32	4.70
104868	KOR	Landrace	Red	3.74	43.56	4.72	6.81	3.19
104914	KOR	Landrace	White	2.01	25.58	0.30	2.14	1.35
105196	KOR	Landrace	Gray	3.30	20.50	2.28	5.92	1.91
105281	KOR	Landrace	Gray	3.83	37.10	4.02	7.65	3.24
105318	KOR	Landrace	Gray	3.92	39.44	4.23	7.83	3.44
105343	KOR	Landrace	Gray	3.34	36.14	3.38	7.13	2.70
105403	KOR	Landrace	Gray	4.17	53.04	5.14	9.58	3.83
105427	KOR	Landrace	Gray	3.97	55.71	4.79	9.10	3.66
105573	KOR	Landrace	Red	3.52	42.10	4.29	6.70	3.23
105591	KOR	Landrace	Red	4.65	61.25	6.44	10.42	4.95
105592	KOR	Landrace	Gray	4.06	47.14	5.39	9.35	3.91
105632	KOR	Landrace	Red	4.52	64.10	6.69	10.11	4.21
105643	KOR	Landrace	Red	4.74	62.66	7.34	10.66	5.04
105698	KOR	Landrace	White	1.52	16.37	0.27	2.02	1.18
105890	KOR	Landrace	White	1.63	19.66	0.23	2.28	1.32
105959	KOR	Landrace	Gray	3.39	26.29	3.06	6.32	2.38
108674	KOR	Landrace	Gray	4.18	60.99	5.97	10.90	4.27
108770	KOR	Landrace	Gray	4.22	32.04	5.73	10.62	4.10
108791	KOR	Landrace	Gray	3.81	29.11	3.17	6.54	2.50
108821	KOR	Landrace	Red	3.91	75.02	4.52	7.28	3.12
108865	KOR	Landrace	Gray	4.56	52.12	5.90	10.02	4.20
108880	KOR	Landrace	Red	2.94	35.43	1.67	7.28	1.84
108932	KOR	Landrace	Gray	4.37	57.51	5.87	9.87	4.16
108935	KOR	Landrace	Red	3.12	36.59	1.36	6.94	2.41
108952	KOR	Landrace	Red	2.97	33.59	1.20	5.62	1.76
109112	KOR	Landrace	Gray	3.42	32.43	2.01	4.90	2.04
110996	KOR	Landrace	Gray	3.40	28.33	2.48	6.34	2.60
111131	KOR	Landrace	Red	2.70	25.30	3.15	6.71	2.29
111146	KOR	Landrace	Red	2.98	29.02	3.29	7.34	2.67
111148	KOR	Landrace	Red	3.86	37.45	5.57	11.54	4.33

	Gei	neral informatior	1		Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg GAE/g)	TSC (mg DE/g)	DPPH (mg AAE/g)	ABTS (mg TE/g)	FRAP (mg AAE/g)
111160	KOR	Landrace	Red	3.09	35.91	3.14	7.30	2.38
111165	KOR	Landrace	Gray	3.63	30.01	2.32	6.74	2.57
112783	KOR	Landrace	Gray	3.92	35.20	2.50	7.14	2.74
112824	KOR	Landrace	Gray	4.27	45.22	3.41	8.95	3.82
112871	KOR	Landrace	White	1.86	20.34	0.27	2.13	1.30
112935	KOR	Landrace	Gray	4.78	46.62	4.03	11.10	4.33
113119	KOR	Landrace	Gray	3.56	35.33	2.50	6.50	2.58
113139	KOR	Landrace	White	1.73	19.60	0.33	2.59	1.46
113215	KOR	Landrace	Red	3.41	30.33	2.95	6.57	3.23
113249	KOR	Landrace	White	2.01	25.15	0.28	2.31	1.52
113259	KOR	Landrace	Gray	4.10	41.33	3.34	9.46	3.54
113383	KOR	Landrace	Red	3.41	35.44	4.18	8.12	3.00
113419	KOR	Landrace	Red	3.34	31.38	3.41	7.54	3.55
113423	KOR	Landrace	White	2.10	24.20	0.33	1.90	1.41
113429	KOR	Landrace	Red	3.51	40.20	4.33	9.23	2.65
113436	KOR	Landrace	Gray	4.09	37.44	2.63	8.13	3.09
113437	KOR	Landrace	Red	3.27	33.44	3.12	7.21	2.97
113455	KOR	Landrace	Gray	3.30	32.84	2.23	6.48	2.38
119917	KOR	Landrace	Gray	3.59	40.43	3.09	8.25	3.24
119922	KOR	Landrace	Gray	4.45	43.76	3.29	8.81	3.54
121444	KOR	Landrace	Gray	4.75	52.47	4.02	8.81	4.51
121459	KOR	Landrace	Gray	5.37	57.97	4.77	10.69	5.21
134952	KOR	Landrace	Gray	4.79	50.23	3.70	7.54	4.11
134972	KOR	Landrace	Gray	4.41	44.63	2.86	5.17	3.31
138107	KOR	Landrace	White	1.91	15.61	0.27	1.50	1.24
138157	KOR	Landrace	White	2.01	26.54	0.29	2.17	1.41
138160	KOR	Landrace	Red	3.28	25.78	2.29	5.36	3.35
142423	JPN	Unknown	Red	3.88	35.81	3.94	8.67	4.66
142424	JPN	Unknown	Red	3.96	48.11	3.90	9.37	5.04
142425	JPN	Unknown	Red	3.97	38.52	3.96	9.20	4.92
142426	JPN	Unknown	Red	3.56	31.63	3.16	7.40	4.09
142427	JPN	Unknown	Red	4.14	38.15	4.49	8.88	5.06
142461	JPN	Unknown	Red	3.75	26.48	3.30	6.33	3.11
142462	JPN	Unknown	Red	4.25	33.25	4.60	7.46	3.81
142463	JPN	Unknown	Red	4.57	46.07	5.81	9.10	4.60
142707	JPN	Unknown	Red	4.01	29.86	4.38	7.54	3.65
158257	KOR	Landrace	Gray	4.77	41.08	3.55	7.16	4.03
158261	KOR	Landrace	Gray	4.71	45.97	3.71	10.16	3.81
162747	KOR	Landrace	White	2.02	28.47	0.28	2.13	1.34
178388	KOR	Landrace	White	2.03	20.61	0.32	2.14	1.50

	Ge	neral informatior	1		Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg GAE/g)	TSC (mg DE/g)	DPPH (mg AAE/g)	ABTS (mg_TE/g)	FRAP (mg AAE/g
178389	KOR	Landrace	Gray	4.82	51.72	4.17	10.74	4.37
178390	KOR	Landrace	White	2.16	28.55	0.52	2.90	1.89
178391	KOR	Landrace	Gray	6.26	60.31	6.11	14.27	5.98
178458	KOR	Landrace	Red	4.29	68.62	5.76	11.37	7.85
178461	KOR	Landrace	Gray	5.14	49.11	4.79	12.34	4.82
178518	KOR	Landrace	Gray	4.62	43.73	3.82	9.51	3.85
178519	KOR	Landrace	Gray	4.76	48.42	3.92	10.08	4.08
178521	KOR	Landrace	Red	3.04	45.37	2.96	6.84	4.75
178527	KOR	Landrace	Red	4.03	56.13	5.41	10.46	7.61
178528	KOR	Landrace	Gray	5.18	51.69	4.14	10.17	4.33
180464	KOR	Landrace	White	2.37	33.85	1.16	3.23	2.56
180468	KOR	Landrace	Red	3.35	55.34	4.44	8.69	6.17
180478	KOR	Landrace	Red	3.68	54.36	4.51	8.82	6.30
180498	KOR	Landrace	Gray	3.08	18.86	1.95	4.73	1.84
180537	KOR	Landrace	Gray	2.99	15.74	1.72	4.20	1.82
180601	KOR	Landrace	Gray	3.72	17.20	2.02	4.93	1.90
180627	KOR	Landrace	Gray	3.96	24.29	3.12	6.98	2.85
180879	KOR	Landrace	White	2.05	23.79	0.33	1.86	1.45
180885	KOR	Landrace	Gray	4.25	29.74	4.03	9.36	3.98
180906	KOR	Landrace	Red	3.81	57.15	4.33	8.28	3.70
180907	KOR	Landrace	Gray	3.86	23.75	2.87	7.19	2.92
180911	KOR	Landrace	White	1.97	25.48	0.32	1.82	1.39
180912	KOR	Landrace	Red	4.73	56.58	4.02	6.90	3.30
180913	KOR	Landrace	Red	3.26	70.61	4.57	9.43	4.10
180915	KOR	Landrace	White	2.08	21.86	0.34	2.27	1.44
180916	KOR	Landrace	Red	3.54	72.13	3.37	6.64	2.84
181898	KOR	Landrace	Red	3.48	49.40	4.05	5.64	2.84
181902	KOR	Landrace	White	2.03	25.75	0.33	2.22	1.45
181958	KOR	Landrace	White	2.04	26.54	0.34	2.59	1.52
186247	KOR	Landrace	Gray	4.59	27.89	3.77	8.61	3.69
186248	KOR	Landrace	Gray	4.71	35.71	4.86	10.24	4.52
186253	KOR	Landrace	Gray	4.42	33.79	4.34	9.78	3.94
186257	KOR	Landrace	Gray	3.70	18.54	1.94	4.94	2.16
186262	KOR	Landrace	White	2.08	28.55	0.34	2.70	1.61
186281	KOR	Landrace	Gray	3.86	26.81	2.43	6.51	3.14
186291	KOR	Landrace	White	1.93	22.95	0.28	2.06	1.40
186294	KOR	Landrace	White	1.86	16.45	0.30	2.05	1.39
186298	KOR	Landrace	Red	4.05	31.33	3.75	8.28	3.93
186299	KOR	Landrace	Red	4.22	38.61	4.23	8.75	4.22
186300	KOR	Landrace	Gray	3.64	25.40	3.20	7.38	3.34

	Ge	neral information			Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg GAE/g)	TSC (mg DE/g)	DPPH (mg_AAE/g)	ABTS (mg_TE/g)	FRAP (mg AAE/g)
186306	KOR	Landrace	Gray	4.64	36.59	4.58	10.20	4.68
186307	KOR	Landrace	Gray	4.29	27.17	3.29	7.55	3.60
186309	KOR	Landrace	Red	5.54	53.56	6.98	13.27	6.26
186310	KOR	Landrace	Red	3.66	31.25	3.30	7.57	3.59
186313	KOR	Landrace	Gray	3.79	26.07	2.75	6.47	3.01
186314	KOR	Landrace	White	1.95	26.59	0.29	2.12	1.42
186322	KOR	Landrace	Gray	5.52	41.10	5.79	12.54	5.97
186324	KOR	Landrace	Gray	4.64	37.23	4.91	11.14	5.06
186325	KOR	Landrace	Gray	4.43	31.40	3.84	8.86	4.07
186326	KOR	Landrace	Gray	4.49	28.23	3.34	7.88	3.54
186327	KOR	Landrace	Gray	4.94	35.98	4.74	10.75	4.70
186328	KOR	Landrace	Gray	4.90	33.40	4.46	9.92	4.46
186330	KOR	Landrace	White	1.93	19.38	0.29	1.06	0.60
186331	KOR	Landrace	Gray	4.24	27.43	3.66	7.92	3.82
186333	KOR	Landrace	White	1.88	23.59	0.27	1.42	0.67
189358	JPN	Unknown	Red	3.53	32.46	4.80	8.19	4.72
189359	JPN	Unknown	Red	3.76	39.57	4.44	8.60	4.53
189360	JPN	Unknown	Red	3.26	24.49	2.82	5.55	3.17
189361	JPN	Unknown	Red	3.59	36.40	3.96	7.05	3.76
189362	JPN	Unknown	Red	3.94	74.76	5.24	9.45	4.94
191131	KOR	Landrace	Gray	4.31	32.54	4.04	8.47	3.83
191175	KOR	Landrace	Red	4.10	42.58	4.24	6.93	3.54
191178	KOR	Landrace	Gray	4.48	32.19	4.38	9.23	4.15
191248	KOR	Landrace	Gray	4.13	46.62	3.02	6.60	3.82
194572	KOR	Landrace	Red	3.32	36.72	3.25	6.46	3.63
194574	KOR	Landrace	Red	3.54	43.17	3.94	7.28	3.96
194575	KOR	Landrace	Red	6.34	114.22	12.84	17.66	13.14
194577	KOR	Landrace	Red	3.25	39.26	3.54	6.35	3.56
195036	KOR	Landrace	Gray	4.05	43.65	3.63	8.25	4.66
195050	KOR	Landrace	Gray	3.52	40.60	2.45	6.35	3.57
195131	KOR	Landrace	White	2.08	22.50	0.36	1.49	0.85
195132	KOR	Landrace	Gray	4.46	43.01	2.48	5.86	3.34
195142	KOR	Landrace	Gray	3.90	70.69	3.03	7.77	4.39
195224	KOR	Landrace	Gray	4.96	77.65	5.13	11.58	6.76
195325	KOR	Landrace	Gray	3.67	70.17	2.88	7.11	4.45
195356	KOR	Landrace	Gray	3.52	70.56	3.35	8.73	5.14
195363	KOR	Landrace	Gray	3.90	73.05	2.60	6.87	4.28
195384	KOR	Landrace	Gray	3.50	47.56	2.35	5.45	3.51
195537	KOR	Landrace	Gray	4.03	82.80	4.17	10.07	6.11
195594	KOR	Landrace	Gray	3.86	80.87	3.79	8.69	5.18

	Ge	neral information	1		Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg GAE/g)	TSC (mg DE/g)	DPPH (mg AAE/g)	ABTS (mg TE/g)	FRAP (mg AAE/g)
196868	KOR	Landrace	Gray	3.82	68.06	3.41	8.34	5.07
196869	KOR	Landrace	Gray	4.16	53.37	2.65	6.30	3.86
196870	KOR	Landrace	Red	4.18	53.53	5.62	8.46	7.32
196871	KOR	Landrace	Red	3.48	46.25	3.82	6.61	5.36
196872	KOR	Landrace	White	1.75	17.50	0.27	1.11	0.59
196894	KOR	Landrace	Red	3.66	42.92	4.21	7.10	6.43
196895	KOR	Landrace	Gray	5.55	57.96	6.47	13.17	8.10
196897	KOR	Landrace	Gray	4.10	77.05	3.71	8.91	5.18
204149	KOR	Landrace	Red	4.03	45.20	4.37	6.66	6.38
204158	KOR	Landrace	White	1.83	16.71	0.29	1.47	0.73
208533	KOR	Landrace	Red	3.40	40.69	3.45	4.88	4.05
208534	KOR	Landrace	Red	3.38	36.41	3.56	4.97	6.32
208535	KOR	Landrace	Red	3.37	34.26	3.14	5.17	5.58
208790	KOR	Landrace	Gray	4.24	82.46	3.55	8.51	5.29
208791	KOR	Landrace	Red	3.89	48.15	4.78	6.89	6.59
208810	KOR	Landrace	Gray	3.63	72.66	3.13	7.76	4.53
208846	KOR	Landrace	Red	3.13	38.39	3.36	3.84	5.27
208890	KOR	Landrace	Red	3.24	32.66	2.23	3.90	3.80
208913	KOR	Landrace	Gray	4.52	89.64	4.64	10.32	6.06
208944	KOR	Landrace	White	1.96	18.17	0.29	1.51	1.12
209326	KOR	Landrace	Gray	3.62	72.62	2.93	7.72	4.71
211854	KOR	Landrace	Gray	4.48	91.36	3.65	9.02	5.45
213272	KOR	Landrace	Gray	3.70	56.59	2.96	6.91	4.03
216285	JPN	Landrace	Gray	5.33	108.24	9.43	16.21	10.84
216286	JPN	Landrace	Gray	8.24	87.61	11.21	14.54	10.50
216289	JPN	Landrace	Gray	5.91	75.35	6.39	10.97	6.60
216290	JPN	Landrace	Gray	4.28	64.65	4.31	10.18	4.31
216291	JPN	Landrace	Gray	5.79	67.10	7.28	10.96	7.55
216295	JPN	Landrace	Gray	3.78	50.84	6.33	12.82	6.85
216298	JPN	Landrace	Gray	6.66	106.16	8.03	14.90	8.37
216299	JPN	Landrace	Gray	6.68	100.91	6.94	14.73	8.09
216300	JPN	Landrace	Gray	6.18	66.62	5.97	14.12	7.18
216303	JPN	Landrace	Gray	7.10	106.44	6.48	14.21	7.51
216305	JPN	Landrace	Gray	7.11	107.21	7.86	16.48	9.67
216306	JPN	Landrace	Gray	7.64	79.42	8.74	16.80	10.09
218257	KOR	Landrace	Gray	3.87	55.58	4.04	9.78	4.74
219512	KOR	Landrace	Gray	3.65	49.18	2.84	6.87	3.16
229635	KOR	Landrace	Gray	4.53	41.47	4.76	11.06	5.36
229704	KOR	Landrace	Gray	3.77	49.13	3.14	7.26	3.50
229706	KOR	Landrace	Gray	4.71	53.51	4.89	11.25	5.50

	Ge	neral information			Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg_GAE/g)	TSC (mg DE/g)	DPPH (mg_AAE/g)	ABTS (mg_TE/g)	FRAP (mg AAE/g)
229962	KOR	Landrace	Gray	4.28	53.81	3.81	9.41	4.41
235359	KOR	Landrace	Gray	4.14	47.46	3.74	9.24	3.93
236188	KOR	Landrace	White	1.98	15.59	0.31	1.38	1.20
236196	KOR	Landrace	Gray	4.28	55.25	3.75	8.70	4.06
236199	KOR	Landrace	White	2.04	19.21	0.28	1.28	1.22
236206	KOR	Landrace	Gray	2.79	32.79	3.74	8.89	4.15
236251	KOR	Landrace	White	2.15	20.13	0.35	1.69	1.27
236259	KOR	Landrace	White	2.08	15.23	0.34	1.05	1.10
236268	KOR	Landrace	Gray	3.52	42.01	2.73	6.72	3.17
236283	KOR	Landrace	Gray	4.25	42.55	3.48	8.82	4.31
236286	KOR	Landrace	Gray	3.81	41.29	3.13	7.57	3.39
236287	KOR	Landrace	Gray	4.03	43.06	2.81	6.31	3.10
236292	KOR	Landrace	White	1.92	18.79	0.32	1.26	1.19
236304	KOR	Landrace	White	2.11	20.34	0.35	1.71	1.32
236596	CHN	Landrace	Gray	4.18	31.28	3.21	7.72	3.59
236597	CHN	Landrace	Gray	3.25	53.73	2.35	4.83	2.56
238022	KOR	Landrace	Gray	3.66	65.80	3.15	6.55	3.66
239918	KOR	Landrace	White	2.15	23.34	0.37	1.93	1.20
239998	KOR	Landrace	Gray	3.68	67.73	3.14	6.94	3.70
239999	KOR	Landrace	Gray	3.69	69.82	3.76	8.31	4.36
240001	KOR	Landrace	Gray	3.69	52.68	2.41	5.14	2.88
240085	KOR	Landrace	Gray	3.87	65.60	3.15	7.09	4.13
240087	KOR	Landrace	Gray	3.69	48.63	3.37	6.90	3.90
240113	KOR	Landrace	Gray	3.79	69.59	3.17	8.16	4.45
240343	KOR	Landrace	Gray	4.90	100.55	4.97	12.09	6.60
240374	KOR	Landrace	Gray	3.67	52.55	1.85	5.21	3.05
242323	KOR	Landrace	Gray	4.10	72.26	3.56	9.28	5.13
242395	KOR	Landrace	Gray	4.03	74.15	2.93	8.29	4.50
252929	KOR	Landrace	Gray	4.40	85.38	4.66	10.57	5.83
252938	KOR	Landrace	Gray	4.03	59.24	2.97	8.49	3.79
252941	KOR	Landrace	Gray	3.88	75.33	3.37	7.89	4.41
263894	KOR	Landrace	Gray	3.83	73.54	3.38	8.05	4.49
263906	KOR	Landrace	Gray	3.82	60.53	3.13	7.98	4.36
280293	CHN	Landrace	Gray	3.99	72.70	3.45	8.29	4.52
280295	CHN	Landrace	Gray	5.16	80.91	4.25	10.09	5.21
280306	CHN	Landrace	Gray	4.05	73.31	3.28	8.46	4.32
280309	CHN	Landrace	Gray	4.36	84.16	3.73	9.38	4.95
283300	KOR	Landrace	Gray	4.85	90.28	4.72	10.63	5.67
286492	KOR	Landrace	Gray	4.12	73.48	2.36	8.80	4.95
290138	KOR	Landrace	Gray	3.77	63.40	1.69	7.15	4.15

	G	eneral information			Metabolite con	ntents and antiox	idant activities	
IT-No.	Origin	Genotype	Seed coat color	TPC (mg GAE/g)	TSC (mg DE/g)	DPPH (mg AAE/g)	ABTS (mg_TE/g)	FRAP (mg AAE/g)
292991	KOR	Landrace	Gray	3.47	28.76	2.38	4.68	2.71
293064	KOR	Landrace	Gray	2.60	47.12	5.34	9.21	5.40
293065	KOR	Landrace	Gray	3.71	41.43	3.94	6.86	4.05
293132	JPN	Landrace	Gray	2.50	19.56	1.13	2.36	1.69
293155	CHN	Landrace	Gray	2.14	21.78	1.81	3.41	2.19
293444	KOR	Landrace	Gray	3.39	45.21	4.70	7.23	4.48
293450	KOR	Unknown	Gray	4.40	55.74	5.50	8.76	5.21
293759	PRK	Unknown	Gray	3.17	47.23	3.68	7.29	4.40
293760	PRK	Unknown	Gray	2.92	37.21	3.20	6.31	3.64
293929	KOR	Landrace	Gray	3.59	35.05	3.68	5.77	3.49
293930	KOR	Landrace	Gray	3.64	45.39	4.15	8.13	4.62
294630	JPN	Breeding line	White	1.85	14.36	0.28	1.60	0.98
294637	JPN	Landrace	Gray	3.24	42.28	3.57	6.82	4.18
296738	KOR	Landrace	Gray	3.08	41.34	3.71	6.72	4.14
296739	KOR	Landrace	Gray	3.59	48.40	4.31	7.91	4.92
318778	KOR	Landrace	Gray	3.14	40.08	3.06	6.10	3.75
		Minimum		1.52	14.36	0.23	1.05	0.59
		Maximum		8.24	114.22	12.84	17.66	13.14
		Mean		3.75	44.27	3.53	7.43	3.84
		SD		1.03	19.72	1.91	3.11	1.80
		CV (%)		27.47	44.54	54.04	41.88	46.79

AAE, ascorbic acid equivalent; ABTS, ABTS^{•+} scavenging activity; CHN: China, CV: Coefficient of variation; DE: Diosgenin equivalent; DPPH, DPPH[•] scavenging activity; FRAP: Ferric reducing antioxidant power; GAE: Gallic acid equivalent; JPN: Japan, KOR: South Korea, PRK: North Korea SD: Standard deviation; TE: Trolox equivalent; TPC: Total phenolic content; TSC: Total saponin content.